

BROWN SEAWEED (*Sargassum ilicifolium*) AS A NUTRITIONAL SUPPLEMENT: EFFECTS ON GROWTH AND HEMATOLOGY IN *Labeo rohita*

Rafia Zulfiqar^{1,*}, Farkhanda Asad^{1,*}, Basim. S. A. Al Sulivany², Saba Naseer¹, Irtisha Kanwal¹, Aiman Nadeem¹, and Muhammad Owais³

¹Department of Zoology, Government College University Faisalabad, Faisalabad, Punjab, Pakistan.

²Department of Biology, College of Science, University of Zakho, Zakho, 42002, Duhok, Kurdistan Region, Iraq.

³Department of Zoology, Emerson University, Multan, Punjab, Pakistan.

*Corresponding author's email: farkhanda.asad@gcuf.edu.pk

Received: 09 Jan. 2025 Accepted: 07 Mar. 2025 Published: 08 Apr. 2025

<https://doi.org/10.25271/sjuoz.2025.13.2.1480>

ABSTRACT:

This experimental trial aimed to evaluate the impact of dietary supplementation with *Sargassum ilicifolium* on the growth performance and hematology of *Labeo rohita*. Fingerlings weighing an average of 2.36g were allocated into the control and three treatment groups (3%, 5%, and 7% BSW), each with triplicate (25 fingerlings/aquarium) for 90 days after acclimatized for two weeks. The results indicated that the highest growth rate ($P < 0.05$) occurred in the T3 treatment group, with a final weight of $5.31 \pm 0.00a$, followed by T2 ($4.84 \pm 0.01b$), and T1 ($4.63 \pm 0.00c$), while the lowest growth was observed in the control group ($4.20 \pm 0.02d$). Maximum weight gain was recorded in T3, which contained the highest level of *Sargassum* supplementation. Other treatments also showed higher weight gain than the control group. Specific growth rate and protein efficiency were also observed to be higher in treatment groups, particularly in T3, than in the control group. Hematological profiles, i.e., red blood cells (RBCs), white blood cells (WBCs), Hemoglobin (Hb), Hematocrit (HCT), blood indices and leukocytes count also showed better results in T3 and other treatments as compared to the control group. These findings suggest a high inclusion of *sargassum* is most effective for promoting the growth and enhancement of the hematology of *Labeo rohita*, and it is an environmentally sustainable way, as it is economical, environmentally friendly, and readily available alternative for aquaculture.

KEYWORDS: *Labeo Rohita*, *Sargassum Ilicifolium*, Growth Performance, Protein Efficiency Ratio, And Hematology.

1. INTRODUCTION

The economic value of algae has gained attention due to their diverse applications in various industries, including aquaculture. In fish feed, algae offer numerous benefits: they provide high-quality protein and essential amino acids, improve feed conversion rates, and contain bioactive compounds with immunomodulatory and anti-inflammatory properties (Veluchamy and Palaniswamy, 2020). Algae are an excellent source of omega-3 fatty acids, which improve nutrition and flesh quality in fish. By adding algae to fish diets, sustainable aquaculture practices can be supported, as this reduces the need for fishmeal and helps lessen the pressure on wild fish populations. In summary, including algae in aquaculture is beneficial as it enhances fish health and promotes sustainability (Liu *et al.*, 2023; Rathod *et al.*, 2025). Algae, as primary producers, are essential to global aquatic ecosystems; they sustain marine food webs and significantly contribute to carbon dioxide fixation and oxygen production (Pereira, L. 2021; Vijayaram *et al.*, 2024). They are utilized for sustenance, pharmaceuticals, cosmetics, and biofuels, among other applications. The seashore of Pakistan is home to around 70 genera and 27 distinct kinds of algae, predominantly seaweeds (Siddiqui *et al.*, 2019).

Based on color, algae fall into three main groups: brown algae (Phaeophyta), green algae (Chlorophyta), and red algae (Rhodophyta) (Shahzad *et al.*, 2023). Among the most important economic algae in Phaeophyta is *Sargassum*. It provides a wide range of development and application prospects in industry,

agriculture, food, medicine, and ecological restoration (Zou *et al.*, 2022), so helping greatly to preserve the marine ecology. There are more than 400 different species of brown seaweed in the genus *Sargassum*, and each has a special ecological use. Common in tropical locations, it produces steroids, polysaccharides, and polyphenols. Bioactive polysaccharides derived from brown seaweeds have gained significant interest among pharmacology and biochemistry researchers. Brown seaweed-derived sulfated polysaccharides have shown anti-inflammatory, antiviral, antioxidant, anti-cancer, and anticoagulant properties (Jyotsna *et al.*, 2021).

Aquaculture, which has been utilized for centuries to produce food, is currently expanding rapidly (Eroldogan *et al.*, 2022). Fisheries provide food for humans and industrial raw materials for national construction, contributing significantly to the national economy and contributing to the country's economy (Hu *et al.*, 2021). Indian major Carp (*Labeo rohita*), commonly known as rohu, is profitable from an economic standpoint, other *Labeo* species could also be cultivated because of their excellent nutritional content, mouthwatering flavor, low economic worth, and high market value. *L. rohita* is one of the fish species that is most often raised in Asia (Khizar *et al.*, 2024).

Sargassum is a desirable addition to the fish diet since it is a bioactive component that is rich in vitamins, minerals, and amino acids (Morais *et al.*, 2020). It stimulates immune responses, fortifies antioxidant defences, and improves growth performance. *Sargassum* improves fish health and resilience to environmental stressors by increasing nutritional absorption, disease resistance, stress tolerance, and digestive enzyme

* Corresponding author

This is an open access under a CC BY-NC-SA 4.0 license (<https://creativecommons.org/licenses/by-nc-sa/4.0/>)

activity. It also improves meat quality and lipid metabolism, rendering fish suitable for human consumption (Saade *et al.*, 2020; Sabzi *et al.*, 2023; Gazali *et al.*, 2024).

This study aims to examine the effects of *S. ilicifolium* on hematological parameters, growth performance, and the immune response of *L. rohita*. Standard fish feed ingredients, which often rely on expensive and scarce resources such as fishmeal and soy protein, may be partially replaced with the sustainable and cost-effective feed additive Sargassum. Given the abundance of sargassum in tropical and subtropical regions, its application in aquaculture reduces dependence on imported feed sources, thus decreasing production costs for fish farmers.

2. MATERIALS AND METHODS

This experimental study evaluated the effect of *S. ilicifolium* supplementation on the growth performance and hematology of *L. rohita*. It was carried out at the Fisheries Laboratory, Department of Zoology, Government College University, Faisalabad, for 90 days.

Collection of *S. ilicifolium*:

Brown seaweed (*S. ilicifolium*) was collected from Manora Beach, Karachi. The seaweed was thoroughly washed under tap water to remove any debris or impurities, followed by another wash with distilled water to ensure complete cleanliness. The cleaned seaweed was dried and ground into a fine powder. This powdered seaweed material was stored at room temperature until further analysis.

Experiment Design:

One control and three experimental diets were prepared with varying amounts of sargassum powder in them. Three hundred fingerlings (2.36g) of *L. rohita* were purchased from Punjab Fish Hatchery, Satyana Road, Faisalabad. Fish were acclimatized for two weeks in the Fish Lab. During acclimatization, fish were fed the control diet of 4% of their body weight once a day. After 15 days, fish were randomly stocked in four aquariums: Control (0%BSW), T1(3%BSW), T2(5%BSW), and T3 (7%BSW), each with triplicates (25 fingerlings per aquarium), arranged using interspersed. Fish were fed with 4 % of their live body weight once a day. Feeding behaviour was also observed in the trial.

Study of Growth Parameters:

Total body length and weight were measured fortnightly over 90 days to track growth performance metrics, according to the approach of Al Sulivany *et al.* (2024). After data collection, fish were stocked back into their respective aquariums. The ethics committee of the Government College University, Faisalabad, Pakistan, approved the animal study protocol for scientific purposes. Approval no. is GCUF/ERC/505.

The following formulas were used to calculate the growth parameters;

$$K = \frac{W \times 100}{L^3}$$

Table 1: Effects of dietary inclusion of *Sargassum ilicifolium* on growth performance of *L. rohita*.

Parameters	Control	T1 (BSW3%)	T2 (BSW5%)	T3 (BSW7%)	P-value
Initial Weight(g)	2.360±0.000a	2.360±0.000a	2.350±0.000a	2.350±0.000a	N. S
Final Weight (g)	4.200±0.030d	4.630±0.030bc	4.840±0.040b	5.310±0.020a	0.0298*
WG (%)	1.570±0.029d	2.270±0.009c	2.490±0.019b	2.960±0.000 a	0.0023**
CF (K)	1.820±0.020d	1.75±0.024c	1.70±0.019b	1.64 ±0.159a	0.0001**
SGR (%)	0.560±0.019d	0.750± 0.009c	0.800± 0.030b	0.900± 0.030a	0.0043**
PER (%)	0.049±0.001d	0.0705±0.002c	0.0779±0.002b	0.092± 0.003a	0.0038**

Values are presented as mean± SE (n = 12 fish from each treatment), WG weight gain, CF condition factor, SGR specific growth rate, and PER protein efficiency ratio

$$WG (g) = \text{Final body weight} - \text{Initial body weight}$$

$$SGR(\%) = \frac{\text{Weight Gain}}{\text{Experimental Days}} \times 100$$

$$PER (\%) = \frac{\text{Weight gain}}{\text{Amount of protein fed}}$$

Analysis of Hematology:

Blood samples were collected from the caudal vein of fish to assess the blood indices. The total number of red blood cells (RBCs) and white blood cells (WBCs) was assessed using a hemocytometer with a validated Neubauer counting chamber (Blaxhall & Daisley, 1973). Hemoglobin concentration estimates were obtained using the cyanomethemoglobin method, as outlined by Wedemeyer & Yastuke (1977). The microhematocrit technique (Brown *et al.*, 1980) was employed to ascertain hematocrit using capillary tubes. The percentage composition of leukocytes was determined based on the identification characters listed (Ivanova, 1983). The subsequent factors were employed for calculation: MCHC (mean corpuscular hemoglobin concentration), MCH (mean corpuscular hemoglobin), and MCV (mean cell volume), utilizing the specified formulas.

$$MCHC (\%) = \frac{Hb}{PCV} \times 100$$

$$MCV (fl) = \frac{PCV}{RBC} \times 100$$

$$MCH (pg) = \frac{Hb}{RBC} \times 100$$

Statistical Analysis:

After the possible outcomes were known, data on growth and hematology were subjected to statistical analysis. ANOVA and ± SE values were computed by using GraphPad ver. 6.0, and the difference between means was calculated by Tukey's HSD test.

3. RESULTS

Growth Parameters:

The effects of *S. ilicifolium* on the growth performance of *L. rohita* are shown in Table 1. After the 90-day trial, Weight gain (WG) and Specific growth rate (SGR) were significantly higher ($P < 0.05$) in groups fed with 5%(T2) and 7% (T3) of *S. ilicifolium* compared to the control, with the highest WG of 2.96 g and SGR of 0.90 observed in the T3. In contrast, the control group exhibited a minimum WG of 1.57 g. The highest Protein Efficiency Ratio (PER) value of 0.092 was recorded in the 7% fed group, significantly higher compared to the control group's PER of 0.049. However, the highest condition factor was observed in the control group, and the lowest was observed in the T3 treatment. Overall, these results suggest the beneficial effects of *S. ilicifolium* on the growth performance of *L. rohita* at higher dietary levels.

Hematological Assay:

Hematological parameters in *L. rohita* improved significantly in fish-fed enriched diets (Table 2). The RBC count was notably higher in fish fed the highest level of 7% (T3) compared to the control group ($P < 0.01$). Hemoglobin values also increased significantly across all treatment groups, with the highest level observed in the T3 (16.5), contrasting with the control group (12.3), where significant differences were noted ($P < 0.01$). WBC counts showed substantial elevation in the T3 (11.4) compared to the control (4.8) ($P < 0.01$). Hematocrit levels

demonstrated a marked increase in the T3 (54.5) versus the control (35.5), with significant differences across all treatments ($P < 0.01$). While MCV, MCH and MCHC values were elevated in the enriched groups, significant differences for MCV were found compared to control ($P < 0.01$). Still, no significant differences in MCH were found between groups ($P > 0.01$). Additionally, the number of blood leukocytes, i.e., Neutrophils, Lymphocytes, Monocytes, Eosinophils and Platelets, was higher in fish fed the enriched diets versus controls, with significant differences particularly noted in the highest inclusion level ($P < 0.01$). Overall, these results indicate that dietary supplementation with *S. ilicifolium* enhances the hematological profile of *L. rohita*, especially at higher inclusion rates

Table 02: Hematological changes of *L. rohita* fed diets enriched with different levels of *Sargassum ilicifolium* (0, 3, 5 and 7%) for 12 weeks

Parameters	Control	T1 (BSW3%)	T2 (BSW5%)	T3 (BSW7%)	P-value
Hemoglobin(g/dl)	12.300±0.200d	12.900±0.100bc	13.00± 0.300b	16.500±0.500a	0.022*
RBC's ($\times 10^6/\mu\text{l}$)	3.750± 0.250d	4.000± 0.300c	4.200±0.400b	5.250± 0.250a	0.0088**
HCT (%)	35.500±0.500d	45.750± 0.750c	48.650±0.350b	54.500±0.500a	0.0001**
MCV (fl)	77.500±0.200d	79.900± 0.100c	88.700± 0.300b	98.500±0.500a	0.0004**
MCH (pg)	25.500±0.500d	29.350± 0.050c	31.100±0.400b	34.500±0.500a	0.0004**
MCHC (%)	30.800±0.200d	33.100±0.400c	35.050±0.750b	37.500±0.500a	0.0029**
WBC's($\times 10^3/\mu\text{L}$)	4.850± 0.150d	8.450± 0.450c	9.850± 0.450b	11.450±0.450a	0.0012**
Neutrophils (%)	40.300± 0.300d	57.900± 0.200c	63.900± 0.100b	73.900± 0.100a	0.0001**
Lymphocytes (%)	24.550± 0.550d	29.500± 0.500c	32.850± 0.150b	39.750± 0.250a	0.0001**
Monocytes (%)	2.300± 0.300d	3.700± 0.300c	4.600± 0.400b	7.250± 0.550a	0.0041**
Eosinophils (%)	1.000± 0.000d	2.000± 0.000bc	2.500± 0.500b	3.600± 0.600a	0.0391*
Platelets($\times 10^3/\mu\text{L}$)	338.250±0.350d	360.000±0.250c	370.000±0.300b	401.900±0.200a	0.0001**

Data are presented as mean \pm SE (n=12 fish from each treatment). Hb: hemoglobin concentration, RBC: red blood cells, WBC: white blood cells, HCT: hematocrit, MCV: mean corpuscular volume, MCH: mean corpuscular hemoglobin, MCHC: mean corpuscular hemoglobin concentration.

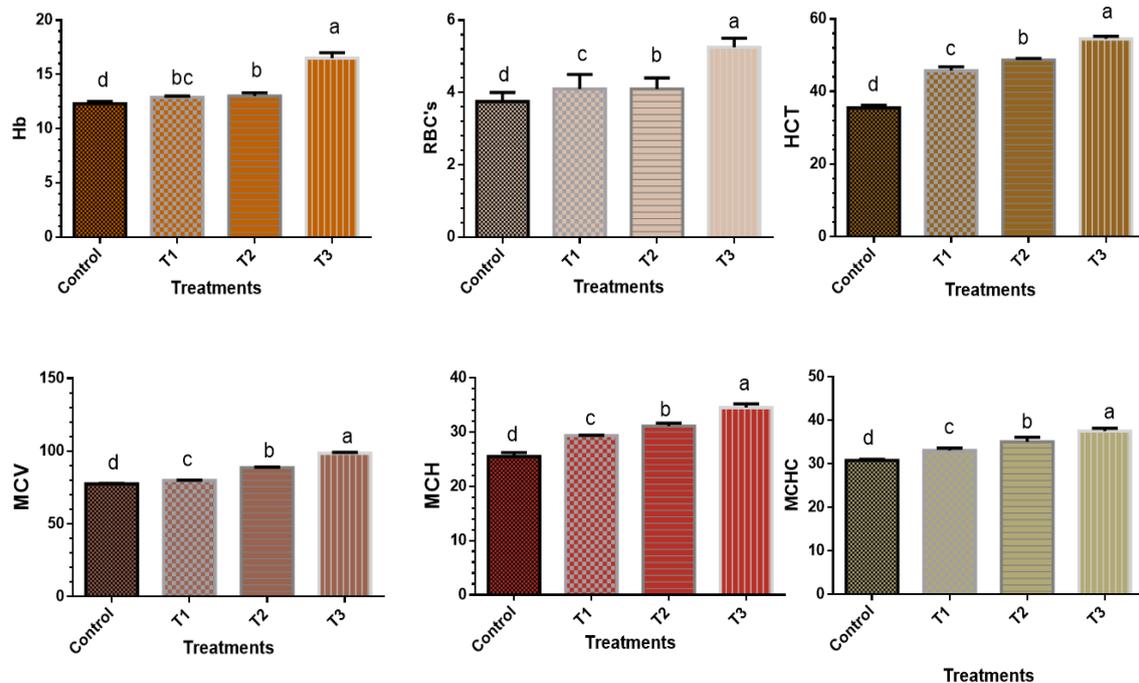


Figure 1: Graph illustration of hemoglobin, red blood cells, hematocrit, MCV, MCH, and MCHC of *L. rohita* reared under control and Treatment groups.

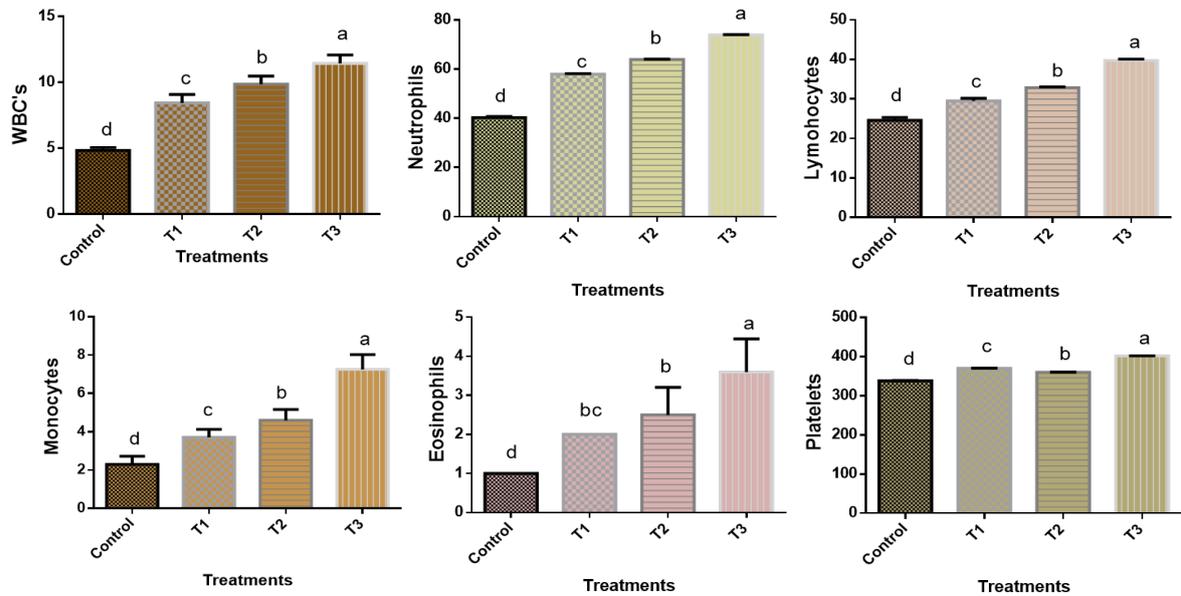


Figure 2: Graph illustration of white blood cells and leukocyte count of *L. rohita* reared under control and Treatment groups

4. DISCUSSION

In the present growth study, the inclusion of *Sargassum* spp. in the feed presented a significant improvement in growth rates associated with the control set. Among inclusion levels, the group with the highest inclusion (7%) showed the best growth performance, with the highest final body weight (FBW), weight gain (WG), and specific growth rate (SGR). The present findings are similar to the studies in which tilapia showed better growth, WG, and SGR when fed with 20 g/kg and 30 g/kg sargassum than the control group (Abdelrhman *et al.*, 2022), Shi *et al.* (2021) stated that yellow catfish, when fed with *Sargassum hornei* extract in feed, enhanced growth and a significant improvement in specific growth rate (SGR) and weight gain (WG) was observed. Yeganeh and Adel (2019) observed that average body weight, total body length, and SGR enhance when supplemented with *S. ilicifolium* extract, particularly at 7.5% inclusion. These results show a favorable association between increased seaweed inclusion and enhanced fish development, with the former showing much superior growth than the latter.

The positive effects observed in the current study align with the findings of Zeynali *et al.* (2020) and Seyed Alhosseini (2023), who found that diets supplemented with *S. ilicifolium* and other seaweed species resulted in improved development performance and feed application. Based on the findings, *Sargassum* spp. Enhances growth in fish as it contains polysaccharides, carbohydrates and bioactive compounds that improve nutritional absorption in fish. Akbary (2019) explored that the highest body weight, condition factor and PER were observed in *Rastrelliger kanagurta* at a 15% inclusion level of *S. ilicifolium*.

In contrast, Shapawi and Zamry (2016) stated that *Sargassum polycystum* in Asian seabass did not improve growth. Similarly, *Palmaria palmata* has no significant effect on the development of Atlantic salmon Wan *et al.* (2016). The inclusion of *A. platensis* in diets of *Oncorhynchus mykiss* did not affect growth performance Teimouri *et al.* (2013). These studies showed that the growth-promoting effects of seaweeds may vary with species-specific nutritional requirements, environmental conditions, seaweed type, and dosage. Plant-based protein sources for fish feed have functional benefits due to their nutritional quality, sustainability, and cost-effectiveness. Asad *et al.*, (2025).

Hematological indices, notably red blood cell count, hemoglobin, and hematocrit values, significantly improved in the

current study when *Sargassum* spp. was added to fish diets at different levels (0, 3%, 5%, and 7%). These results are consistent with earlier research by Sabzi *et al.* (2023), which found that feeding fish species like *L. calcarifer*, *Cirrhinus mrigala*, and *Oreochromis niloticus* diets supplemented with seaweed significantly increased their RBC, Hb, and Hct levels.

With increasing dietary inclusion of *Sargassum* spp., RBC values increased significantly. This was especially true for the 7% inclusion group, which showed the largest significant increases in RBCs, Hb, and Hct compared with the control. The developments imply that *Sargassum* spp. may contain bioactive compounds, such as polysaccharides and antioxidants, which, by enhancing oxygen delivery and immune function, could improve fish blood parameters and overall health.

Furthermore, the notable increase in neutrophils and WBC in diets supplemented with *Sargassum* spp. in fish-eating diets is a crucial indicator of immunological activation, which is consistent with the results of Yeganeh and Adel (2019), who also observed elevated WBC levels. Given that the body uses these cells to fight infections, the observed increase in neutrophils in our study suggests a stronger immunological response. The 7% *Sargassum* spp. The diet showed a notable increase in neutrophils, thus substantiating the hypothesis that *Sargassum* spp. can enhance fish immune systems.

Several investigations have found the opposite of these encouraging findings. A study conducted by Yang *et al.* (2014) found that fucoidan, a chemical derived from brown seaweed, had no significant effect on the red and white blood cell counts or hematocrit levels in yellow catfish.

In conclusion, the present study confirms that dietary inclusion of *Sargassum* spp. enhances hematological indices in fish, particularly RBC, Hb, Hct, and WBC levels. These improvements in blood demonstrate that *Sargassum* spp. contributes to better overall physiological status and immune function in fish. However, some studies have also reported neutral or negative effects of seaweeds.

CONCLUSION

The findings of this study demonstrated that the growth performance and immunological response of *Labeo rohita* were significantly improved in the treatment groups with dietary inclusion of *Sargassum ilicifolium* compared with the control group. Compared with the treatments, the best results were observed with 7% sargassum inclusion (T3), followed by 5%

inclusion (T2), and the least, but still better than control, results were observed with 3% sargassum inclusion (T1).

Acknowledgements

The authors extend their sincere appreciation to the Department of Zoology, Government College University Faisalabad, Faisalabad, Punjab, Pakistan, for providing the academic environment and resources that facilitated the successful publication of this work.

Statements and Declarations:

Conflict of interest: Not applicable

Consent to Participate: All authors have agreed to submit the article to this journal

Consent to Publish: All authors agree to publish in this journal

Funding: No funding

Data availability statement: On request

REFERENCES

- Abdelrhman, A. M., Ashour, M., Al-Zahaby, M. A., Sharawy, Z. Z., Nazmi, H., Zaki, M. A., Van & Doan, H. (2022). Effect of polysaccharides derived from brown macroalgae *Sargassum dentifolium* on growth performance, serum biochemical, digestive histology and enzyme activity of hybrid red tilapia. *Aquaculture Reports*, 25, 101212. <https://doi.org/10.1016/j.aqrep.2022.101212>
- Akbary, P. (2019). The effect of supplementation of algae (*Sargassum ilicifolium*) on growth, feed and body chemical composition of the Indian mackerel (*Rastrelliger kanagurta*). *Experimental animal Biology*, 8(2), 143-151. <https://doi.org/10.30473/eab.2019.6212>
- Al Sulivany, B. S., Hassan, N. E., & Mhammad, H. A. (2024). Influence of Dietary Protein Content on Growth Performance, Feed Efficiency, Condition Factor, and Length-Weight Relationship in *Cyprinus carpio* during the Summer Season. *Egyptian Journal of Aquatic Biology & Fisheries*, 28(2). <https://doi.org/10.21608/ejabf.2024.349722>
- Asad, F., Al Sulivany, B. S., Hassan, H. U., Nadeem, A., Rohani, M. F., Owais, M., ... & Arai, T. (2025). Evaluating the Differential effect of growth and health parameters on *Oreochromis niloticus* and *Cirrhinus mrigala* under Difference rice protein Concentration. *Egyptian Journal of Aquatic Research*. <https://doi.org/10.1016/j.ejar.2024.12.002>
- Blaxhall, P. C., & Daisley, K. W. (1973). Routine haematological methods for use with fish blood. *Journal of Fish Biology*, 5(6), 771-781. <https://doi.org/10.1111/j.1095-8649.1973.tb04510.x>
- Brown, S., Caro, J., Erslev, A. J., & Murray, T. G. (1980). Spontaneous increase in erythropoietin and hematocrit value associated with transient liver enzyme abnormalities in an anephric patient undergoing hemodialysis. *The American Journal of Medicine*, 68(2), 280-284. [https://doi.org/10.1016/0002-9343\(80\)90367-8](https://doi.org/10.1016/0002-9343(80)90367-8)
- Eroldogan, O. T., Glencross, B., Novoveska, L., Gaudencio, S. P., Rinkevich, B., Varese, G. C., & Rotter, A. (2023). From the sea to aquafeed: A perspective overview. *Reviews in Aquaculture*, 15(3), 1028-1057. <https://doi.org/10.1111/raq.12740>
- Gazali, M., Effendi, I., Husni, A., Nurjanah, N., Wahyuni, S., & Kurniawan, R. (2024). Sargassum sp. extract improve hematological profile of tilapia fish (*Oreochromis niloticus*). *F1000Research*, 12(293). <https://doi.org/10.12688/f1000research.128819.4>
- Hu, F., Zhong, H., Wu, C., Wang, S., Guo, Z., Tao, M., & Liu, S. (2021). Development of fisheries in China. *Reproduction and Breeding*, 1(1), 64–79. <https://doi.org/10.1016/j.repbre.2021.03.003>
- Jyotsna Golden, C. D., Koehn, J. Z., Shepon, A., Passarelli, S., Free, C. M., Viana, D. F., & Fluet-Chouinard, E. (2021). Aquatic Foods to Nourish Nations. *Nature*, 598(7880), 315–320. <https://doi.org/10.1038/s41586-021-03917-1>
- Ivanova, N.T. (1983). Atlas of fish blood cells. Comparative morphology and classification of fish blood cells. Moscow: Light and Food Industry, 182 p.
- Khizar, A., Fatima, M., Khan, N., & Rashid, M. A. (2024). Effects of phytase inclusion in diets containing rice protein concentrate (RPC) on the nutrient digestibility, growth, and chemical characteristics of rohu (*Labeo rohita*). *Plos one*, 19(5), e0302859. <https://doi.org/10.1371/journal.pone.0302859>
- Liu, Y., Cao, L., Cheung, W. W., & Sumaila, U. R. (2023). Global estimates of suitable areas for marine algae farming. *Environmental Research Letters*, 18(6), 064028. <https://doi.org/10.1088/1748-9326/acd398>
- Morais, T., Inácio, A., Coutinho, T., Ministro, M., Cotas, J., Pereira, L., & Bahcevandziev, K. (2020). Seaweed potential in the animal feed: A review. *Journal of Marine Science and Engineering*, 8(8), 559. <https://doi.org/10.3390/jmse8080559>
- Pereira, L. (2021). Macroalgae. *Encyclopedia*, 1(1), 177-188. <https://doi.org/10.3390/encyclopedia1010017>
- Rathod, S. G., Choudhari, A. N., & Mantri, V. A. (2025). A Global Bibliometric Analysis of Seaweed Biodiversity, Endemic Taxa, and Conservation (1992–2023). *Phycology*, 5(1), 1. <https://doi.org/10.3390/phycolgy5010001>
- Saade, E., Sulaeman, H., & Jannah, N. (2020). The effects of seaweed, Sargassum sp. meal dosages in the artificial diet on growth, feed intake, feed efficiency, protein efficiency ratio, and nutritional body composition of Rabbitfish, *Siganus guttatus*. Paper presented at the IOP Conference Series: *Earth and Environmental Science*, 564(1), 12049. <https://doi.org/10.1088/1755-1315/564/1/012049>
- Sabzi, E., Mohammadiarm, H., & Salati, A. P. (2023). Synergistic effects of *Sargassum vulgare* extract and lipid levels on growth performance, blood biochemical indices, immunological competence, and antioxidant capacity in juvenile common carp (*Cyprinus carpio*). *Aquaculture Reports*, 33, 101829. <https://doi.org/10.1016/j.aqrep.2023.101829>
- Seyedalhosseini, S. H., Salati, A. P., Mozanzadeh, M. T., Parrish, C. C., & Shahriari, A. (2023). Effects of dietary seaweeds (*Gracilaria* spp. and *Sargassum* spp.) on growth, feed utilization, and resistance to acute hypoxia stress in juvenile Asian seabass (*Lates calcarifer*). *Aquaculture Reports*, 31, 101663. <https://doi.org/10.1016/j.aqrep.2023.101663>
- Shahzad, S. M. (2023). Seaweeds as potential product for Pakistan's blue economy: A review. *Pakistan Journal of Weed Science Research*, 29(3), 156-163. <https://dx.doi.org/10.17582/journal.PJWSR/2023/29.3.156.163>
- Shapawi, R., & Zamry, A. A. (2016). Response of Asian seabass, *Lates calcarifer* juvenile fed with different seaweed-based diets. *Journal of Applied Animal Research*, 44(1), 121-125. <https://doi.org/10.1080/09712119.2015.1021805>
- Shi, Q., Wang, J., Qin, C., Yu, C., Wang, S., & Jia, J. (2021). Growth performance, serum biochemical parameters, immune parameters and hepatic antioxidant status of yellow catfish *Pelteobagrus fulvidraco* supplemented with *Sargassum horneri* hot-water extract. *Aquaculture Reports*, 21, 100839. <https://doi.org/10.1016/j.aqrep.2021.100839>

- Siddiqui, M. D., Zaidi, A. Z., & Abdullah, M. (2019). Performance evaluation of newly proposed seaweed enhancing index (SEI). *Remote Sensing*, 11(12), 1434. <https://doi.org/10.3390/rs11121434>
- Teimouri, M., Amirkolaie, A. K., & Yeganeh, S. (2013). The effects of *Spirulina platensis* meal as a feed supplement on growth performance and pigmentation of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, 396, 14-19. <https://doi.org/10.1016/j.aquaculture.2013.02.009>
- Veluchamy, C., & Palaniswamy, R. (2020). A review on marine algae and its applications. *Asian Journal of Pharmaceutical and Clinical Research*, 13(3), 21-27. <https://doi.org/10.22159/ajpcr.2020.v13i3.36130>
- Vijayaram, S., Ringø, E., Ghafarifarsani, H., Hoseinifar, S. H., Ahani, S., & Chou, C. C. (2024). Use of Algae in Aquaculture: A Review. *Fishes*, 9(2), 63. <https://doi.org/10.3390/fishes9020063>
- Wan, A. H., Soler-Vila, A., O'Keeffe, D., Casburn, P., Fitzgerald, R., & Johnson, M. P. (2016). The inclusion of *Palmaria palmata* macroalgae in Atlantic salmon (*Salmo salar*) diets: effects on growth, haematology, immunity and liver function. *Journal of Applied Phycology*, 28, 3091-3100. <https://doi.org/10.1007/s10811-016-0821-8>
- Wedemeyer, G., & Yastuke, T. (1977). A method for determining blood hemoglobin concentration in fish. *The Progressive Fish-Culturist*, 39(2), 127-130.
- Yang, Y., Zhang, X., Zhang, Y., Liao, Y., & Liu, L. (2014). Effects of fucoidan on growth performance and blood parameters of yellow catfish (*Pelteobagrus fulvidraco*). *Fish Physiology and Biochemistry*, 40(4), 1221-1230. [10.1016/j.fsi.2014.09.003](https://doi.org/10.1016/j.fsi.2014.09.003)
- Yeganeh, S., & Adel, M. (2019). Effects of dietary algae (*Sargassum ilicifolium*) as immunomodulator and growth promoter of juvenile great sturgeon (*Huso huso* Linnaeus, 1758). *Journal of Applied Phycology*, 31(3), 2093-2102. <https://doi.org/10.1007/s10811-018-1673-1>
- Zeynali, M., Nafisi Bahabadi, M., Morshedi, V., Ghasemi, A., & Torfi Mozanadeh, M. (2020). Replacement of dietary fishmeal with *Sargassum ilicifolium* meal on growth, innate immunity and immune gene mRNA transcript abundance in *Lates calcarifer* juveniles. *Aquaculture Nutrition*, 26(5), 1657-1668. <https://doi.org/10.1111/anu.13111>
- Zou, X., Meng, T., Yao, D., Chen, Z., Zhu, J., Mu, D., & Bao, S. (2022). Benthic *Sargassum* composition and community characteristics in the intertidal zone of Hainan Island, China. *Marine Biology Research*, 18(9-10), 555-565. <https://doi.org/10.1080/17451000.2023.2174262>