

NEUROTOXICITY OF URANYL ACETATE IN MALE RATS

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Abstract

The present investigation included the neurotoxic effect of different doses of uranyl acetate in male rats. Routine light and electron microscopic techniques were used in this study. The histological studies of cerebellum and cerebrum showed a significant increase of died and decrease of healthy pyramidal cells in the 3rd layer of cerebrum and Purkinje cells in the cerebellum. This may indicate the passage of uranium across the blood brain barrier or disruption of this barrier due to uranium exposure..

Keywords: Cerebellum, Cerebrum, Purkinje cells, uranyl acetate

Introduction

Uranium, a naturally occurring heavy metal, is found in all soils, water, and rocks (Craft *et al.*,2004).The release of uranium into the environment presents a threat to human and ecological health in many parts of the world (Murray *et al.*,2002). Its extensive use in the nuclear cycle and for military applications has focused attention on its potential health effects (Carriere *et al.*,2004). It penetrates through lung tissue and by ingestion to the blood stream and can be stored in the liver, kidney ,or other tissues for years (Patolka *et al.*,2004;Carriere *et al.*, 2005; Li *et al.* .,2005; Paquet *et al.*,2006).Other important possible targets of uranium exposure are bone (Kurttio *et al.*,2005) and brain in which it is transferred across the blood brain barrier (BBB) (Lemercier *et al.*,2003; Lestaeve *et al.*, 2005; Paquet *et al.*,2006).Uranium was found to induce behavioral effects in rats(Belles *et al.*,2005).

Uranium given as uranyl acetate added to the mice chow showed highest accumulation in the brain compared to liver and kidney(Ozmen and Yurekli 1998). Genetic and histological effect of uranium in fish brain was detected by Lerebourset *et al.*, (2010).

The aim of the current work was to study the neurotoxic effect of uranium as uranyl acetate in rat.

Material and Methods

Thirty male albino rats (*Rattus norvegicus*) weighing (230 - 250 gm) and 8-10 weeks old were used in the present study. The animals were bred and housed in plastic cages (56 x 39 x 19 cm) bedded with

wooden chips in animal house / Biology Dept. /College of Science /Salahaddin University-Erbil. Temperature was set as (22 ± 2 C).Regular 12-hours diurnal cycles were kept using an automated light-switching devise. The animals were supplied with water and standard diet *ad libitum*.

The studied rats were divided into 5 groups (six in each). Four groups were given daily oral aqueous doses (5 , 10,20,and 40 mg/kg b.w.) of uranyl acetate dehydrate (UAD) (Fluka AG , Buche SG)for 30 days against a control group which only received water.

Light microscopy

Samples of cerebellum and cerebrum were removed from the anesthetized animals . The samples were fixed in formol saline. All samples were processed for light microscopy by embedding in paraffin after dehydration and clearing. Four micrometers thick sections were stained by hematoxylin and eosin(Bancroft *et al.*,1977).

Electron microscopy

Tissue samples were fixed in 2.5% glutaraldehyde in 0.1 cacodylate buffer pH 7.2-7.4 for 24 h ,postfixed in 1% osmium tetroxide for 2 h , dehydrated in ethanol, cleared in propylene oxide , and embedded in araldite mixture. Plastic blocks were sectioned by ultramicrotome (Riechert co.) into 1µm and 600-900Å° thick sections for light and electron microscopy respectively. The ultrathin sections were mounted on copper grids. One micrometer thick sections were stained by toluidine blue ,while the ultrathin sections were stained by uranyl acetate and lead citrate(Veeramachaneni *et al.*, 1993) . Jeol TEM120 II transmission

electron microscopy (at Kuwait University/Medical College) at 80Kv was used for examining and photographing the ultrathin sections.

For counting dying and healthy cells per millimeter square, five sections through each of the cerebellum and cerebrum were employed. All the measurements were performed using a Lietz microscope equipped with standardized square. The area of this square was calculated under oil immersion lens using stage micrometer and it was equal to 0.005184 mm². The total number of pyramidal cells of the 3rd layer of cerebral cortex and the Purkinje cells of cerebellum (healthy and died cells) were counted per standardized square and transformed to the number per square millimeter of counted region. The above procedure of counting cells was achieved according to the counting procedure of (Abdel-Rahman and Zeki 1992).

Results

The total number of pyramidal cells of the second and third layer of cerebral cortex and Purkinje cells of the middle layer of cerebellum/mm² showed significant decrease ($p < 0.05$) in the UA treated group compared to control. On the other hand, the number of died pyramidal and Purkinje cells/mm² was significantly increased ($p < 0.05$) compared to control in a dose dependent manner (Fig 1 and 2 respectively and Table 1). The mode of cell death of both cells was apoptosis-like mode of cell death (Fig 3) in which the cells appeared shrunken and nucleus condensed or fragmented. Electron microscopic figures revealed different degeneration levels of pyramidal and Purkinje cells (Fig 4 & Fig 5 respectively). Some cells appeared died according to the programmed cell death in which the cell appeared shrunken and many apoptotic vacuoles were seen around the died cell (Fig 5).

Table(1): Effect of different doses of uranyl acetate on brain

Doses mg/kg/d	Total pyramidal cells/mm ²	Died pyramidal cells/mm ²	Total Purkinje cells/mm ²	Died Purkinje cells/mm ²
0	2228.33±3.59 ^a	35.33±2.91 ^a	689.00±4.90 ^a	4.33±0.13 ^a
5	2182.00±3.73 ^b	364.67±3.02 ^b	630.50±1.26 ^b	60.67±1.76 ^b
10	2109.00±5.91 ^c	465.20±5.20 ^c	578.80±2.81 ^c	90.40±1.21 ^c
20	1921.67±3.66 ^d	514.67±3.92 ^d	505.83±2.68 ^d	156.00±3.63 ^d
40	1730.29±3.97 ^e	704.29±4.06 ^e	464.14±3.79 ^e	201.86±2.57 ^e

Values in vertical columns followed by a different letter are significantly different at the 1% level. N=6 animals per group.

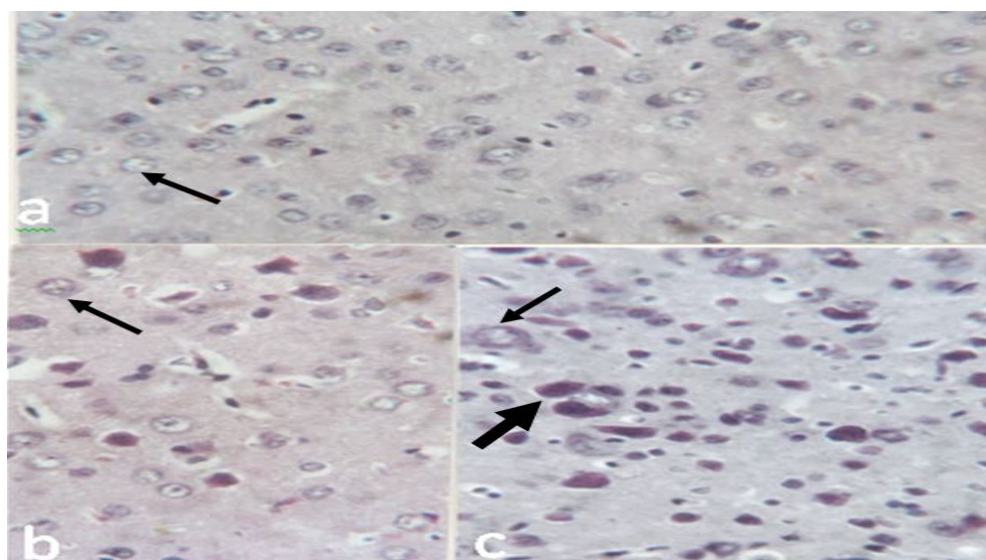


Fig 1: Plastic sections of rat cerebral cortex showing pyramidal cells in the 3rd layer of the cortex, a) control section, b) 10mg UA/kg/d treated, c) 40mg UA/kg/d treated rats (All mag. were 1000X), small arrow=healthy cells, large arrows =died cells

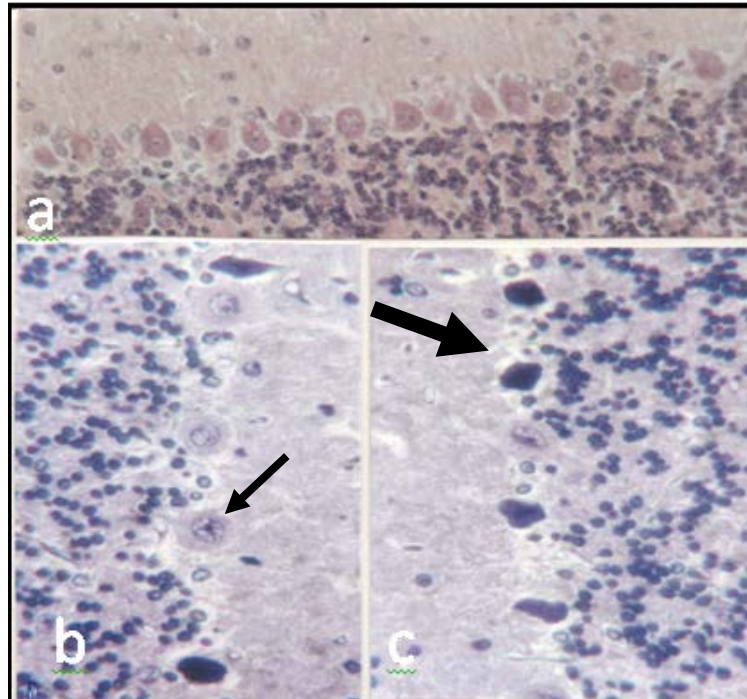


Fig 2 : Cerebellar cortex of UA treated rats a) Paraffin section of control b) 5mg UA/kg/d treated group c) 40mg UA/kg/d treated group. All mag. were 400X. Notice healthy Purkinje cells (small arrows) and died Purkinje cells (large arrows)

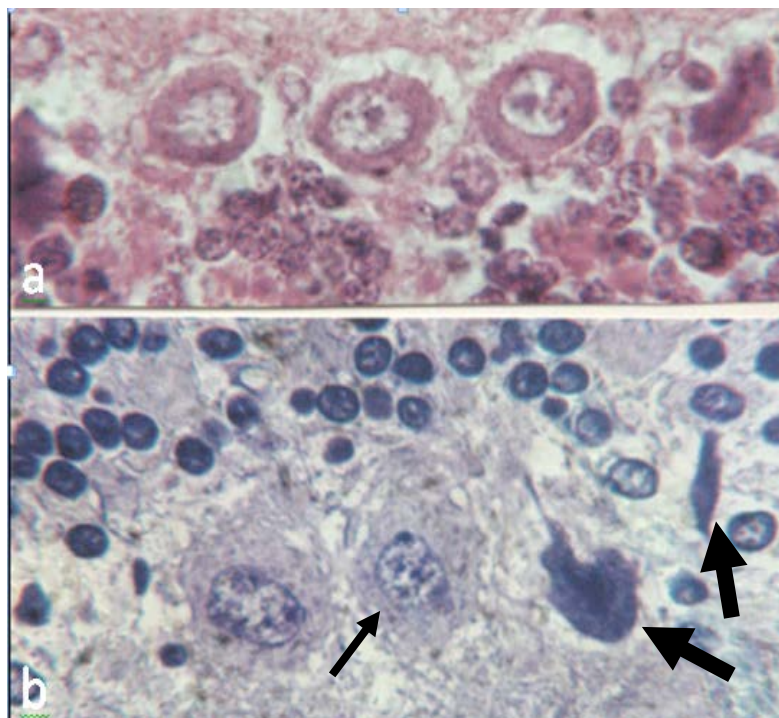


Fig 3: High magnification of Purkinje cells in the a) control, b) 10mg UA/kg/d treated rats showing healthy (small arrows) and died Purkinje cells (large arrows), both are 1000X.

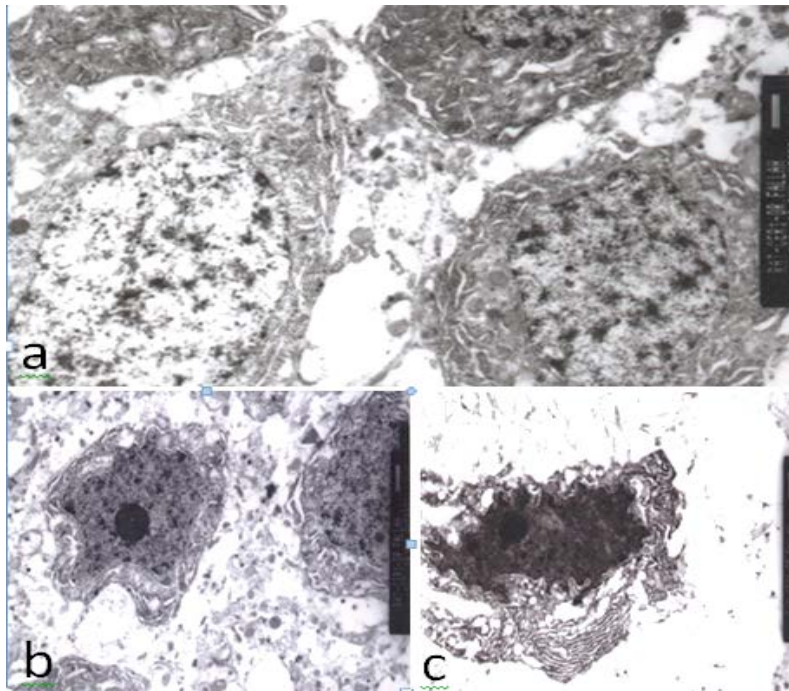


Fig 4:Electron micrographs of pyramidal cells in the UA treatedrat cerebral cortex showing a)normal appearance bar=1μm , b)bar=1μm & c) degenerated shrunken cell .bar=1μm

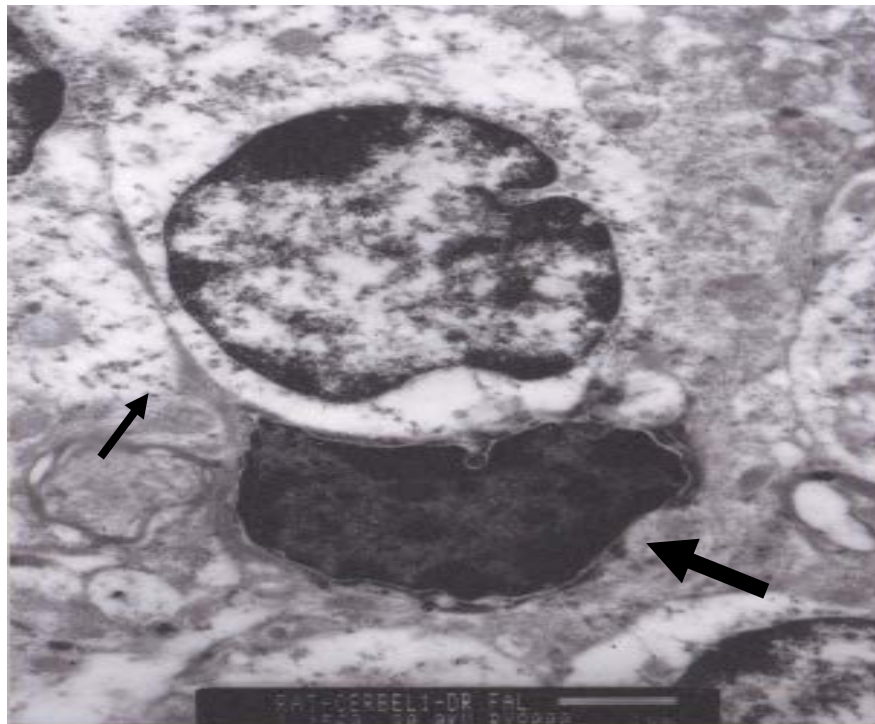


Fig 5: Electron micrograph of UA treated rat cerebellar cortex showing the healthy (small arrow)and died (large arrow) Purkinje cells bar=5μm.

Discussion

The present work detected a significant decrease of healthy pyramidal and Purkinje cells/mm² which may be due to the degenerative effect of uranium on these cells. This degenerative effect was reflected by the significant increase of died neuronal cells. The mode of cell death of these cells was apoptotic-like cell death. These effects indicated that this metal may cross the blood-brain barrier (BBB). This crossing was suggested by several studies which confirmed the accumulation of uranium in the brain of animals after uranium ingestion through natural uranium ingestion (Lemerrier *et al.*, 2003), uranyl acetate uptake (Ozmen and Yurekli, 1998; Barber *et al.*, 2005; Briner and Murray 2005), uranyl nitrate ingestion (Paquet *et al.*, 2006) implanted uranium pellets (Pellmar *et al.*, 1999) or injected depleted uranium (DU) (Lestaeve *et al.*, 2005). The latter paper referred to a dose dependent increase fashion of uranium in the rat brain with the number of implanted DU pellets and this may explain our results with respect to the dose dependent decrease of pyramidal and Purkinje cells/mm² and increase of died cells. The latter effect may also be due to the oxidative stress caused by uranyl acetate treatment. Such oxidative stress through free radicals production has been reported in mammals and other vertebrates such as fishes (Gagnaire *et al.*, 2013). Oxidative stress can induce neuronal cell death in a variety of circumstances (Halliwell, 1992). On the other hand, Abou-Donia *et al.*, (2002) suggested that DU could induce an increase in the generation of nitric oxide in the cortex of the animals treated with 0.1 mg/kg uranyl acetate. The latter compound produced in the CNS is a highly reactive species that has been implicated in a variety of neurodegenerative diseases (Gobbe *et al.*, 1997; Squadrito and Pryor, 1998; Bogdan, 2001). Furthermore, it is also possible that the long-term health effects following exposure with uranyl acetate may be a consequence of changes in the BBB permeability (Abou-Donia *et al.*, 2002). Briner and Murray (2005) detected behavioral effect and lipid oxidation in the brain of rats related to the amount of DU exposure. Lipid oxidation may alter ionic conductance, cell membrane fluidity, or a number of other

cellular functions (Schaich 1992). As a conclusion, uranyl acetate may cause several dose dependent neurotoxic effects as a result of neurocytes degeneration in the exposed rats.

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التأثيرات السمية العصبية لخلايا اليورانيوم في ذكور الجرذان

الخلاصة

تضمن البحث الحالي دراسة التأثير العصبي للجرعات المختلفة لخلايا اليورانيوم في ذكور الجرذان. واستخدمت طرائق تقليدية مجهرية ضوئية والكثرونية لدراسة الخلايا العصبية في المخ والمخيخ. أظهرت الدراسات النسيجية زيادة ملحوظة في عدد الخلايا العصبية الهرمية الميتة في الطبقة الثانية والثالثة لقشرة المخ في المجاميع المعاملة بخلايا اليورانيوم، بينما لوحظ انخفاض في عدد الخلايا السليمة من النوع نفسه. ولوحظت النتيجة نفسها فيما يخص خلايا بركنجي الموجودة في المخيخ وكانت تلك التغييرات في كلا النوعين من الخلايا طردية مع زيادة جرعة خلايا اليورانيوم. قد تشير هذه النتيجة الى اختراق عنصر اليورانيوم الحاجز الدموي الدماغي.

كاريگهري ژههراوی دهماری سرکهی یورانیوم له جرجی سپیدا

پوخته

ئهم تویژینه وهویه لیکۆلینه وهی کاریگهري دهماری ژهه می جۆراوجۆری سرکهی یورانیوم له جرجی نیر. لهه تویژینه وهیه دا، ریگای پشکنینی شانەزانی میکروسکۆبی رووناکي وئەله کترۆنی به کارهینرا بۆ دیراسه تکرندی دهمارخانه کانی مێشک و مێشکۆله. تویژینه وه که ده ریجست که ژماره ی دهماره خانه مردوه کانی تویژی مێشک له زیادبووندا بوو لهو گروپانه ی که سرکهی یورانیومی پیدرا، به لآم خانه ساغه کان به ره وه که مپوون چوون. هه مان ئەنجام بۆ خانه کانی پرکنجی مێشکۆله دهست کهوت. ئەو گۆرنکاریانه له گه ل زیاد کردنی ژههه کانی سرکهی یورانیوم ته ریب بوو. ئەم ئەنجامه له وانیه یه ئەوه ده رنجات که توخمی یورانیوم به به ره بهستی خوین و مێشک تیپه ر بیّت.