

## Review article

# ECOLOGICAL EFFECTS AND ENVIRONMENTAL FATE OF SILVER NANOPARTICLES IN SOIL AND IN THE WATER ECOSYSTEM: A REVIEW

Asma Umer<sup>1</sup>, Basim. S. A. Al Sulivany<sup>2,3</sup>, Riffat Yasin<sup>4</sup>, Muhammad Jamshed<sup>1,\*</sup>, Maqbool Ahmad<sup>1</sup>, Inayat Ullah Malik<sup>5</sup>, Muhammad Tauqeer Riaz<sup>1</sup>, Muhammad Shoaib Akhtar<sup>5</sup>, Muhammad Luqman Tauhidi<sup>5</sup>, Muhammad Owais<sup>5</sup>, Khizar Samiullah<sup>5</sup>, Rana Mehroz Fazal<sup>5</sup>

<sup>1</sup>Department of Chemistry, Ghazi University, Dera Ghazi Khan, Pakistan.

<sup>2</sup>Department of Biology, College of Science, University of Zakho, Zakho, 42002, Duhok, Kurdistan Region, Iraq.

<sup>3</sup>Anesthesia Department, college of health sciences, Cihan University-Duhok, Iraq.

<sup>4</sup>Department of Zoology, Education University, Dera Ghazi Khan, 03221, Punjab, Pakistan.

<sup>5</sup>Department of Zoology, Ghazi University, Dera Ghazi Khan, Pakistan.

\*Corresponding author. E-mail: [mjamshed@gudgk.edu.pk](mailto:mjamshed@gudgk.edu.pk) (Tel.: +923335106562)

## ABSTRACT

Received:  
17, Nov, 2025

Silver nanoparticles, which are appreciated due to their increased antibacterial, catalytic and conductive functions, are commonly employed in medical kits, cloths, cosmetics, and water filters, but their release through wastewater, biosolids, and runoff is highly dangerous as there is no regulation of their emission and transformation, such as sulfidation, aggregation, and dissolution. In soil ecosystems, AgNPs react with pH, redox conditions, organic matter, and clay, and sulfidation to Ag<sub>2</sub>S causes short-term bioavailability to decrease but increases persistence; they disrupt microbial communities, inhibit nitrogen-fixing bacteria (e.g., Rhizobium), mycorrhizal fungi, and enzyme activities, decrease soil fertility, nutrient cycling, and plant-microbe symbiosis and cause oxidative stress in earthworms. Aquatic systems facilitate AgNP disaggregation, sedimentation, and ion release driven by organic matter and ions and cause toxicity at all trophic levels: algae experience the inhibition of photosynthesis and ROS damage, zooplankton feeding problems, and fish experience bioaccumulation, neurotoxicity and reproductive problems. Ag<sup>+</sup> ion release leading to protein/DNA damage, Oxidative stress due to ROS, membrane peroxidation, quorum sensing disruption and systemic changes in stress, detoxification and metabolism pathways confirmed by omics is a subset of the toxicity mechanisms. Though the water body information is plentiful, soil research is still very limited; gaps still exist in long-term low dose field effects and co-contamination. The research in the future recommends mesocosm/field testing, model dynamic transformation, and the safer design of nanoparticles to guide the risk analysis and sustainable management.

Accepted:  
09, Dec, 2025

Published:  
08, Jan, 2026

**KEYWORDS:** Silver nanoparticles; Environmental fate; Ecotoxicology; Aquatic ecosystems; Soil ecosystems.

## 1. INTRODUCTION

Nanotechnology has enabled the production of nanoparticles of silver (AgNPs) that exhibit enhanced physical, chemical, and biological characteristics that exceed those of bulk silver (Wang *et al.*, 2018). Nano-sized materials are used to enhance catalytic activity, antibacterial properties, and electrical conduction, thereby expanding their applications in medical kits, antimicrobial protection, wound dressings, textiles, water filters, cosmetics, and packaging materials (Ghobashy *et al.*, 2021). These applications utilize the microbial membrane-disrupting properties of AgNPs against microbial cells, making them

invaluable in infection control and hygiene management (Sharma *et al.*, 2019). Despite the rapid expansion of AgNP production due to the expanding consumer market for nano-enabled products, environmental risk assessments and regulatory measures have not kept pace (Khan *et al.*, 2023). Therefore, although the usefulness remains unquestionable, widespread, unregulated use of AgNPs requires a simultaneous assessment of their ecological and long-term environmental consequences (Dang *et al.*, 2021).

AgNPs enter the environment mainly via wastewater discharge, leaching out of landfills, and the surface runoff of agricultural soils treated with nano-enabled agrochemicals

Access this article online

<https://doi.org/10.25271/sjuz.2026.14.1.1838>

Science Journal of University of Zakho  
Vol. 14, No. 01, pp. 30 –38, January-2026



Printed ISSN 2663-628X;  
Electronic ISSN 2663-6298

This is an open access under a CC BY-NC-SA 4.0 license  
(<https://creativecommons.org/licenses/by-nc-sa/4.0/>)

(Dodds *et al.*, 2021). Once liberated, silver nanoparticles are too small to settle due to their colloidal nature and may remain suspended in water bodies or within permeable soil matrices, where they may undergo intricate transformation processes (León-Silva *et al.*, 2016). Research indicated that traditional wastewater treatment facilities cannot effectively eliminate nanoparticles, leading to their buildup in sewage sludge or release into natural water sources (McGee, 2020). AgNPs present in bio solids are often used as fertilizers in agricultural environments, where they have direct contact with soil biota/crops (Arienzo & Ferrara, 2022; Asad *et al.*, 2025).

Once in environmental compartments, AgNPs do not remain chemically stable; their properties are altered due to pH, redox potential, organic matter, and ionic strength (Khan & Akram, 2020). Nanoparticle contamination is a major concern for soil ecosystems, which are microbial diverse systems, and are indispensable to global nutrient cycling (Tangaa *et al.*, 2016). AgNPs exposure may also inhibit symbiosis plant-microbe by inhibiting nitrogen-fixing bacteria and mycorrhizal fungi (Rajput *et al.*, 2020). Research has reported a lowering of enzymatic activity, low microbial respiration, and changes in the turnover of carbon and nitrogen in the soil after exposure to AgNP. Such disturbances reduce not only the fertility of the soil but also above-ground productivity and biodiversity (Jahan *et al.*, 2017). Additionally, plant roots can absorb and transfer AgNPs to plant tissues above ground, where they may proceed into the food chain may causing further damage to herbivores and even human beings (Du *et al.*, 2018). Species that are detritivores, such as earthworms, are key in soil aeration and decomposition of organic matter, and evidence has been found that there are oxidative stress, stunted growth, and reproductive abnormalities in these organisms when they ingest AgNPs, thus indicating larger repercussions at the population level (Kalantzi *et al.*, 2019).



**Figure 1:** Summary of the industrial uses, the environmental exposure pathways, and biological hazards of AgNPs.

AgNPs in soil undergo sulfidation, aggregation, and complexation with organic matter, which affect microbial activity, plant health, and nutrient cycling. By contrast, aquatic systems facilitate dissolution, photoreduction, and ion release,

which contribute to oxidative stress and bioaccumulation and can alter aquatic food webs. Although data on aquatic toxicology are sufficient, studies on soil are relatively narrow in scope and duration. It is essential to recognize that transformation pathways and vulnerabilities to the ecological environment vary between these settings, underscoring the necessity to tailor risk assessment to each specific situation and adopt sustainable methods for nanoparticle management (Padhye *et al.*, 2023).

#### PRISMA-Guided Systematic Review:

An extensive and systematic literature review was done according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) (<https://prisma-statement.org/>) protocol to provide methodological soundness and transparency. The search protocol targeted three of the largest academic databases, Web of Science (<https://www.webofscience.com/>), Scopus (<https://www.scopus.com/>), and PubMed (<https://pubchem.ncbi.nlm.nih.gov/>), to identify as many interdisciplinary investigations as possible on the environmental fate of AgNPs. Research works that focused on environmental exposure pathways, trophic relationships, bacterial responses, and toxicological effects in non-human beings were given a higher priority (Kurwadkar *et al.*, 2015). Articles were, in turn, not included when they paid attention to medical applications, synthetic protocols, or in vitro mammalian toxicology, and did not carry the ecological element. The removal of duplicate records and non-peer-reviewed (opinion pieces, conference abstracts, etc.) sources were also provided through additional filtering (Buffet *et al.*, 2014). By using these criteria of inclusion and exclusion, 162 peer-reviewed articles published between the years 2010 and 2025 were found appropriate to be subject to a deeper level of analysis. The combination of these studies covers a variety of geographic areas, experimental geometry, and environmental conditions, which provides a solid background on assessing trends in pattern AgNP transformation, exposure pathways, and ecological risk exposure in terrestrial and aquatic environments (Akhter *et al.*, 2024).

#### Ag NPs Environmental Fate:

AgNPs' fate in the environment depends on their size, surface coatings, and shape, and exogenous environmental factors, including pH, redox conditions, ionic strengths, and organic matter contents (Li *et al.*, 2020). In the environment, AgNPs have been shown to exhibit dynamic transformations involving aggregation, dissolution to Ag<sup>+</sup> ions, sulfidation, and complexation with natural ligands (Furtado *et al.*, 2015). AgNPs persist in soil, affecting microbes and nutrient cycling, while in water, they remain bioavailable and toxic to aquatic life. While soil acts as a terminal sink, aquatic systems remain continuously polluted because wastewater treatment cannot fully remove AgNPs (Zhang *et al.*, 2019).

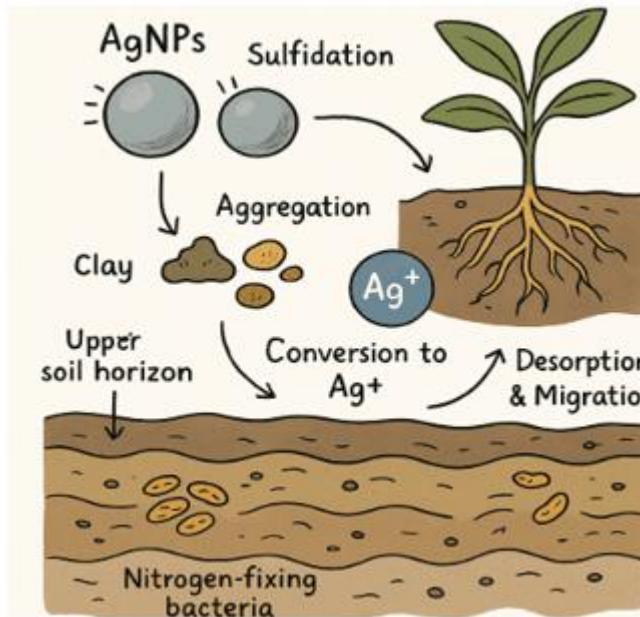
#### AgNP in Soil Ecosystems:

AgNPs interact dynamically in soils, influenced by factors like pH, redox potential, organic matter, cation exchange capacity, and texture. Introduced via biosolids, agricultural inputs, or leachate, AgNPs can undergo sulfidation in sulfur-rich or anoxic environments, forming Ag<sub>2</sub>S (Eivazi & Afrasiabi, 2018). Moreover, AgNPs spontaneously aggregate with clay minerals and become bound to humic and fulvic acids, which contribute to the surface adsorption of AgNPs, mostly at the topsoil roll (Grün *et al.*, 2019). However, changes in pH, rainfall, or flooding can release AgNPs or ions, enabling vertical movement into subsoil, particularly in coarse-textured soils like sandy loams (Yang *et al.*, 2017).

In addition to their geochemical activity, AgNPs produce a dramatic impact on the biological activity of soil microbiota and plant-microbe interaction (Kulikova, 2021). Oxidative dissolving

of AgNPs leads to the liberation of silver ions ( $\text{Ag}^{1+}$ ), which is an extremely active form that inhibits crucial essential activities in the beneficial soil organisms (Peyrot et al., 2014). The effects of  $\text{Ag}^{1+}$  can be particularly toxic to nitrogen-fixing bacteria like Rhizobium and Azotobacter, with the presence of  $\text{Ag}^{1+}$  shown to affect colony-forming units, activity of associated enzymes (e.g., nitrogenize), and symbiotic root nodulation (Cao et al., 2017). Such disruptions not only decrease nitrogen availability to plants, but also undermine larger soil fertility and ecosystem services (de Oca-Vásquez., et al., 2020). Moreover, AgNPs exposure has been known to decrease microbial carbon biomass and phosphatase responses, another way to express deteriorating microbial-mediated nutrient cycling (Zhang et al., 2020). Therefore, transformation processes can lead to changes in AgNP mobility, but the concomitant formation of detrimental species of silver introduces an ongoing ecological risk to the landscape food webs and crop production continuity (López-Mondéjar et al., 2020).

Figure 2 illustrates AgNPs in soils undergo sulfidation to form  $\text{Ag}_2\text{S}$ , reducing short-term bioavailability while increasing long-term persistence (Zhang et al., 2020).

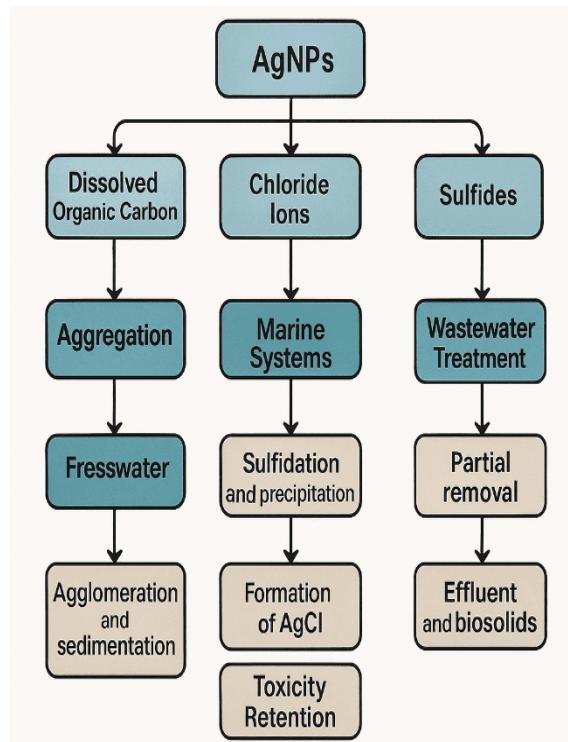


**Figure 2:** Diagrammatic representation of transformation, mobility, and the effect of AgNP in the soil ecosystem with important connections to soil components and microbial communities.

## Aquatic Environments:

Aquatic environment forms a highly heterogeneous and chemically active environment where the fate and behavior of AgNPs are determined via a wide-ranging transformation mechanism. AgNPs are usually disaggregated in freshwater because of their interaction with divalent cations and natural organic matter, and such contacts decrease colloidal stability (Ellis et al., 2016). The results of this aggregation frequently sediment out of the water column and are laid down in sediments, particularly in lakes and slow-flowing rivers (Ottoni et al., 2020).

Figure 3 shows how AgNPs interact with dissolved organic carbon in water, leading to aggregation, agglomeration, and sedimentation, particularly in freshwater bodies (Sohn et al., 2015).



**Figure 3:** Major environmental routes, conversions, and fate of silver nanoparticles (AgNPs) in waters, which determined dissolved organic matter, chloride ions, and sulfides.

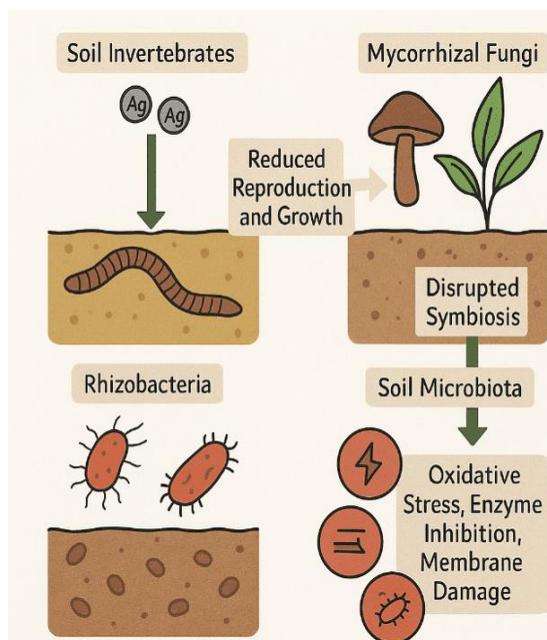
## Ecotoxicological Impacts:

AgNPs exert ecotoxicological effects on several trophic levels through the destabilization of physiologic and biochemical mechanisms of affected organisms (Noga et al., 2023). Their small, reactive particles enter cells, causing oxidative stress, membrane damage, enzyme inhibition, and genotoxicity (Singh et al., 2022). AgNPs disrupt nutrient cycling, plant-microbe interactions, and microbial biomass by impairing nitrogen fixation and other key soil enzyme activities (Tonczyk et al., 2025). AgNPs and  $\text{Ag}^{1+}$  disrupt algal photosynthesis, alter zooplankton feeding, and bioaccumulate in fish, causing inflammation, neurotoxicity, and reproductive failure (Aslam et al., 2024).

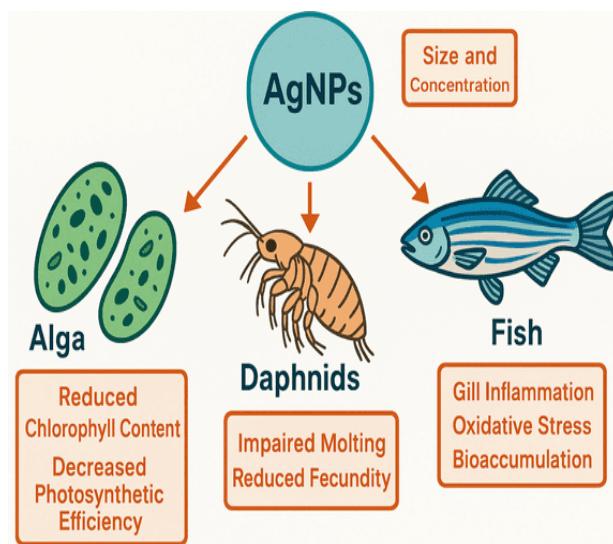
## Organisms in soil:

Soil invertebrates, such as earthworms (*Eisenia fetida*) and springtails (*Folsomia candida*), are key bioindicators of soil health (Courtois, 2020). Environmentally relevant doses of AgNPs reduce growth, reproduction, and alter behavior (Oktarina, 2017). AgNPs enter soil invertebrates via integument penetration or ingestion, localizing in gut tissues and disrupting physiology (He et al., 2016). They induce ROS production, causing oxidative stress, mitochondrial dysfunction, DNA damage, and protein impairment. In earthworms, this leads to gut epithelial disruption and lipid peroxidation (Grün et al., 2017).

In addition to the invertebrates, AgNPs, notably, the plant-root interaction microbes, also affect the beneficial microorganisms in the soil negatively. Mycorrhizal fungi, which aid the uptake of water and nutrients, and rhizobacteria, which aid in nitrogen fixation and the production of phytohormones, are very sensitive to exposure to nanoparticles (Tortella et al., 2020). The presence of AgNPs is especially highlighted in a multi-tiered ecotoxicological effect on soil biota, as shown in Figure 4.



**Figure 4:** Mechanistic diagram of the effects of AgNP on soil organisms, including effects on invertebrates, mycorrhizal fungi, rhizobacteria, and microbial community.



**Figure 5:** AgNP Toxicity mechanisms in aquatic biota approaches, representation in a schematic view with the point of **Figure 6:** Mechanism of toxicity.

**Table 1:** Describing the main ways in which AgNPs have cell toxicity, like, release of ions, the production of ROS, the disruption of membranes, and the disruption of microbes.

Mechanism Title	Description	Biological Implications	Ref.
<b>Ag<sup>+</sup> Ion Release Causing Protein and DNA Damage</b>	AgNPs undergo partial dissolution under oxidative or acidic conditions, releasing Ag <sup>+</sup> ions. These ions bind to thiol groups in cysteine-rich proteins, leading to denaturation, and interact with DNA, causing strand breaks.	Genotoxicity, impaired protein function, apoptosis, and mutation risk	(De Matteis et al., 2015)

### Aquatic life:

Aquatic organisms exhibit a pronounced susceptibility to the toxic effects of AgNPs, with toxicity pathways strongly influenced by particle size, concentration, and the mode of exposure (Chaachouay *et al.*, 2024). *Chlorella vulgaris* and similar algae are primary producers in aquatic systems, making them among the first things to be exposed to AgNPs (Kusi & Maier, 2022).

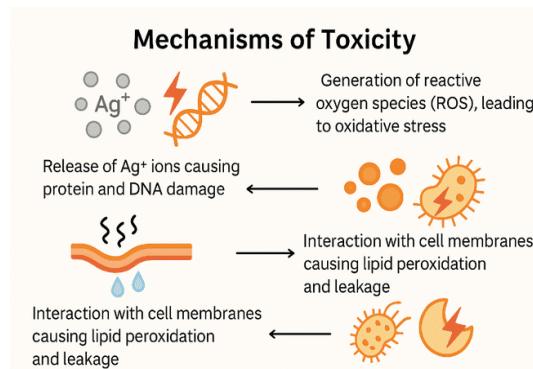
Exposed AgNPs adsorb to the algal cell surface, or cell wall, into which they enter, interfering with chloroplast action and thylakoid membrane integrity. This leads to the reduction of chlorophyll levels and a malfunction of photosystem II, with such a detrimental effect on photosynthetic efficiency (Lapresta-Fernández *et al.*, 2012; Al Sulivany *et al.*, 2025). Reactive oxygen species (ROS) generation increases cellular stress in algae, leading to oxidative damage and reduced photosynthetic efficiency, thus weakening the aquatic food (Kwok *et al.*, 2012). Additionally, nanoparticle surface coatings such as citrate, polyethylene glycol, or polyvinylpyrrolidone strongly influence AgNP dissolution and cellular uptake, modulating algal toxicity (Kang *et al.*, 2023).

Figure 5 presents the specific organism adverse pathways that occur because of the AgNP exposure to the aquatic life.

defining organism-specific impacts, as well as the effects of particle size and surface chemistry.

### Mechanisms of Toxicity:

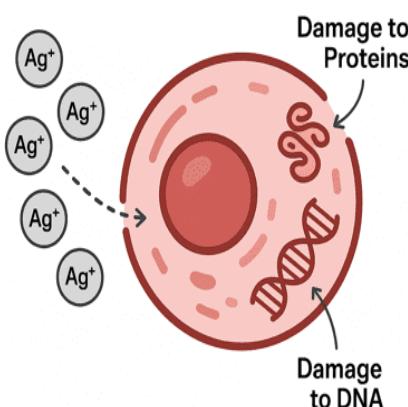
The fundamental AgNP cellular toxicity mechanisms can be described using Figure 6. Ag<sup>+</sup> release interferes with the structure of the protein and causes DNA to fragment, and the production of ROS causes oxidative stress and pervasive damage to the cell (De Matteis *et al.*, 2015).



<b>ROS Generation and Oxidative Stress</b>	AgNPs catalyze the formation of ROS such as superoxide, hydrogen peroxide, and hydroxyl radicals. These overwhelm cellular antioxidants, leading to lipid peroxidation, protein oxidation, and mitochondrial dysfunction.	Redox imbalance, inflammation, mitochondrial damage, chronic toxicity (Flores-Lopez et al., 2019)
<b>Membrane Interaction Causing Lipid Peroxidation and Leakage</b>	AgNPs adhere to phospholipid membranes, disrupting their structure. Lipid peroxidation leads to membrane destabilization, ion leakage, and osmotic imbalance, often triggering necrotic or apoptotic cell death.	Cell lysis, impaired homeostasis, necrosis, or apoptosis initiation (Paciorek et al., 2020)
<b>Disruption of Microbial Quorum Sensing and Enzymatic Pathways</b>	AgNPs disrupt bacterial communication by degrading or blocking signaling molecules, suppressing biofilm formation and gene expression. Ag <sup>+</sup> ions inhibit key enzymes involved in energy and nitrogen metabolism.	Suppressed microbial activity, biofilm inhibition, nutrient cycle collapse (Gomez-Gomez et al., 2019)
<b>Omics-Based Evidence of Systemic Disruption</b>	Omics studies reveal widespread gene expression changes linked to stress, detoxification, and inflammation. Proteomics and metabolomics confirm disruption in cytoskeletal proteins and energy/lipid metabolism.	Whole-organism dysfunction, prolonged stress responses, impaired metabolic regulation (Abdelkader et al., 2023)

#### Ag ions leaking into Cells, resulting in Damage to Proteins and DNA:

Silver ions (Ag<sup>+</sup>) release due to one of the most basic mechanisms of AgNP toxicity has highly characteristic biochemical activities. AgNPs can also easily transform into a partial dissolution form to release Ag<sup>+</sup>-agents, especially in oxidation conditions or acidic environments. The effects of silver ion (Ag<sup>+</sup>) infiltration into cells are illustrated in Figure 7. Once inside, Ag<sup>+</sup> binds to thiol-containing proteins and nucleic acids, disrupting their structural integrity (Li et al., 2017).



**Figure 7:** Oxidative stress is linked to intracellular Ag<sup>+</sup> infiltration, which damages proteins and DNA as presented.

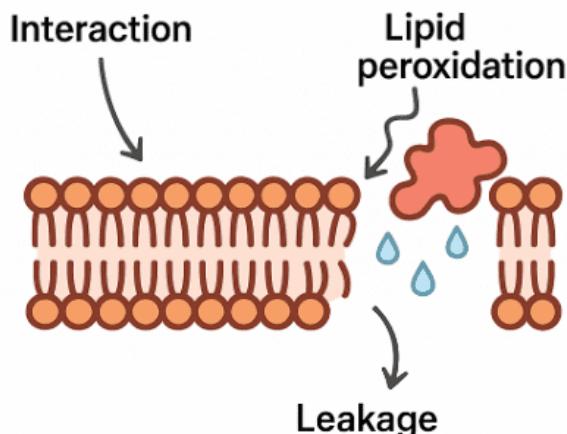
#### Reactive Oxygen Species production and oxidative stress:

It is also known that AgNPs can catalyze the generation of ROS, including superoxide radicals, hydrogen peroxide, and hydroxyl radicals. This occurs by redox cycling at the nanoparticle surface or as secondary responses of cells to Ag<sup>+</sup>. The production of ROS throws the intercellular redox balance out of control and overburdens antioxidant countermeasures like glutathione, superoxide dismutase, and catalase (Saini et al., 2016). This leads to widespread cellular damage due to oxidative stress, such as the peroxidation of lipids, carbonylation of proteins, and oxidative changes in nucleic acids (He, 2013).

#### Interaction of AgNPs with Cell Membranes leading to Lipid Peroxidation and Loss of Membrane Integrity:

Another important mechanism of AgNPs is direct interactions with cell membranes. Due to their small size and high surface area, AgNPs readily interact with phospholipid

membranes, increasing permeability and losing selective ion gradients (Ahmed et al., 2018; Paciorek et al., 2020). These interactions promote lipid peroxidation and membrane destabilization, leading to osmotic imbalance, enzyme inhibition, and ultimately necrotic or apoptotic cell death (Nayak et al., 2016). The interaction of nanoparticles or reactive oxygen species with the lipid bilayer of the cell membrane induces peroxidation of the lipid (Figure 8).

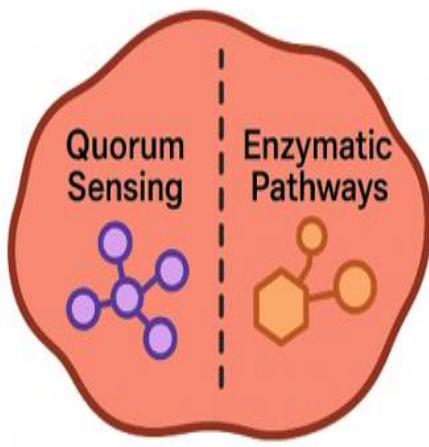


**Figure 8:** Lipid peroxidation produces disruption of the cell membranes, resulting in structural damage and intracellular leakage.

#### Influence on Bacterial quorum sensing and enzymatic pathways:

AgNPs also affect metabolism and intercellular communication, besides compromising the cellular structure in the microbial system (Awadelkareem et al., 2023). The chemical communication chain known as quorum sensing (QS), through which bacteria synchronize group behavior like biofilm production, aggressiveness, and sharing of resources, is highly susceptible to the interference of AgNP (Shah et al., 2019). AgNPs may have the ability to suppress QS-controlled gene expression by breaking down signal molecules (e.g., acyl homoserine lactones) or attaching to receptor sites (Wolska et al., 2017). Furthermore, the presence of released Ag<sup>+</sup> ions into the nanoparticles distorts important enzyme reactions, including the ATP production process, nitrogen fixation (e.g., nitrogenase activity), and dehydrogenase activity (Qadeer et al., 2024). The effect of this disturbance ultimately reduces bacterial growth and survival (Khusro et al., 2023).

Figure 9 reveals the spatial and functional division in bacterial cells between quorum-sensing mechanisms and enzymatic pathways (Peixoto et al., 2022).

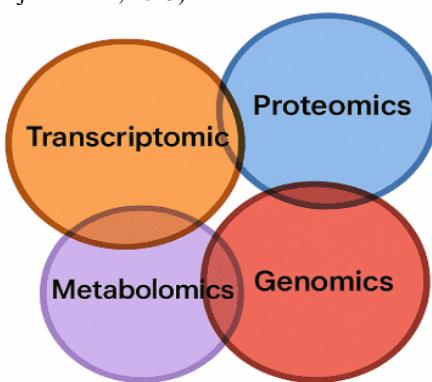


**Figure 9:** The division of quorum-sensing processes and enzymatic processes means that the behavior and metabolism of bacteria can be carefully regulated.

#### Evidence of Systemic Disruption by Omics:

Recent developments in high-throughput omics technologies like transcriptomics, proteomics, and metabolomics have offered molecular-scale insights into systems-level responses of organisms to AgNPs exposure (Aragoneses-Cazorla et al., 2022). Research indicates that transcriptome profiles are altered, involving oxidative stress reactions, metal detoxification (e.g., metallothioneins), inflammation, and apoptotic signaling pathways (Anh et al., 2023). As an example, upregulation of stress-related genes (HSP70, GPX) could reflect successful efforts in adapting cells, whereas downregulation of genes related to mitochondrial respiration and cell division reflects intrinsic toxicity (Gavin, 2016). The proteomic studies demonstrate that the expression of cytoskeleton and membrane proteins has changed, whereas in the metabolomics, alterations were observed in amino acid, lipid, and energy metabolism (Qi et al., 2024). Such multiple-layered perturbations highlight a systemic cellular response to AgNP exposure, indicating that the toxicity is not limited to a few specific pathways but coordinated stress response in a biological system as a whole (Li et al., 2024).

The combined study of omics layers, genomics, transcriptomics, proteomics, and metabolomics is offered in Figure 10 to identify systemic biological perturbations (Andrejević et al., 2023).



**Figure 10:** Multi-omics levels unmask systemic molecular perturbations in terms of their genomic, transcriptomic, proteomic, and metabolomic aspects.

#### Future Research Directions:

To fill research gaps, AgNP studies should extend beyond the lab to mesocosm and field experiments, which simulate natural conditions and capture ecological interactions such as microbial responses, nutrient cycling, and trophic effect (Shaikh et al., 2021). Such extensive validation plans will not only prove the environmental applicability of the previous results but will also help in elaborating regulatory guidance and risk assessment methods, which are based on the ecological reality (Jangid et al., 2024). AgNPs is the examination of nanoparticle transformation processes occurring under the variable environmental conditions, particularly to the variation of redox potential, seasonal changes, and microbial activity (Xiao et al., 2019). In dynamic ecosystems, AgNPs rarely remain in their original form; they continuously undergo oxidation, sulfidation, aggregation, or corona formation, often driven by microbial activity or environmental factors such as temperature fluctuations and photoperiod changes (Rajkuberan et al., 2015). AgNP transformations can alter surface charge, reactivity, and bioavailability, affecting toxicity and environmental persistence (Chutrakulwong et al., 2024; Vivekanandhan et al., 2012). Laboratory microcosms combined with omics tools can clarify these changes and guide safer-by-design strategies (Yadav & Sahu, 2024).

#### CONCLUSION

AgNPs significantly impact both soil and aquatic ecosystems due to their small size, high surface activity, and ion release. They disrupt microbial diversity and enzyme functionality critical for nutrient cycling, affecting soil fertility and plant health. AgNP longevity is influenced by their interactions with soil components, which affect their mobility and bioavailability. In aquatic environments, AgNPs cause oxidative stress and alter trophic relationships, with toxicity often linked to the release of Ag<sup>+</sup> ions. Bioaccumulation raises concerns for long-term ecosystem health, especially in nutrient-rich or slow-flowing waters. Current research reveals knowledge gaps related to long-term low-concentration exposure and interactions with co-contaminants, while most studies focus on short-term, single-species models. Lack of standardized methodologies hampers comparison and risk assessment efforts. Future approaches should combine laboratory, mesocosm, and field studies, utilizing advanced modeling and omics technologies to enhance understanding of AgNP effects, alongside prioritizing environmentally friendly synthesis and design strategies.

#### Acknowledgment:

The authors acknowledge the Department of Chemistry and the Department of Zoology, Ghazi University, Dera Ghazi Khan, Pakistan, for providing academic support and facilities necessary for completing this review. The authors are also thankful to all colleagues who provided valuable suggestions during the preparation of the manuscript.

#### Ethical Statement:

This article is a review based on previously published literature. It does not involve any experiments on humans or animals, and therefore, ethical approval was not required.

#### Author Contributions:

Conceptualization, A.U., M.J., and B.S.A.A.S.; literature review, A.U., R.Y., and M.J.; data analysis, M.A., I.U.M., and M.T.R.; writing original draft preparation, A.U. and I.U.M.; writing review and editing, R.M.F. and B.S.A.A.S.; supervision, B.S.A.A.S. All authors have read and agreed to the published version of the manuscript.

**Funding:**

This research received no external funding.

**REFERENCES**

Abdelkader, Y., Perez-Davalos, L., LeDuc, R., Zahedi, R. P., & Labouta, H. I. (2023). Omics approaches for the assessment of biological responses to nanoparticles. *Adv Drug Deliv Rev*, 200, 114992. <https://doi.org/10.1016/j.addr.2023.11499210.1016/j.addr.2023.114992>

Al Sulivany, B. S. A., Omar, I. . . , Yousif , A., & Owais, M. . (2025). Effects of Dietary Green Microalgae (*Chlorella vulgaris*) and Iron Nanoparticles on Biochemical, Enzymatic, and Tissue Health in *Cyprinus carpio*. *Journal of Aquaculture Science*, 10(2), 98–108. <https://doi.org/10.20473/joas.v10i2.74314>

Akhter, M. S., Rahman, M. A., Ripon, R. K., Mubarak, M., Akter, M., Mahbub, S., Al Mamun, F., & Sikder, M. T. (2024). A systematic review on green synthesis of silver nanoparticles using plants extract and their bio-medical applications. *Heliyon*, 10(11). <https://doi.org/10.1049/nbt2.12078> 10.1049/nbt2.12078

Anh, N. H., Min, Y. J., Thi My Nhung, T., Long, N. P., Han, S., Kim, S. J., Jung, C. W., Yoon, Y. C., Kang, Y. P., Park, S. K., & Kwon, S. W. (2023). Unveiling potentially convergent key events related to adverse outcome pathways induced by silver nanoparticles via cross-species omics-scale analysis. *J Hazard Mater*, 459, 132208. <https://doi.org/10.1016/j.jhazmat.2023.132208>

Aragoneses-Cazorla, G., Buendia-Nacarino, M. P., Mena, M. L., & Luque-Garcia, J. L. (2022). A Multi-Omics Approach to Evaluate the Toxicity Mechanisms Associated with Silver Nanoparticles Exposure. *Nanomaterials* (Basel), 12(10), 1762. <https://doi.org/10.3390/nano12101762>

Arienzo, M., & Ferrara, L. (2022). Environmental fate of metal nanoparticles in estuarine environments. *Water*, 14(8), 1297. <https://doi.org/10.3390/w14081297>

Asad, F., Nadeem, A., Naseer, S., Ashraf, A., Sulivany, B., & Jamal, R. (2025). Toxic and synergistic effects of micro-nanoplastics with radioactive contaminants on aquaculture: Their occurrence, distribution, role as vectors, detection and removal strategies. *International Aquatic Research*, 17(2), 95-116. doi: [10.22034/iar.2025.2008924.1739](https://doi.org/10.22034/iar.2025.2008924.1739)

Aslam, H., Umar, A., Nusrat, N., Mansour, M., Ullah, A., Honey, S., Sohail, M. J., Abbas, M., Aslam, M. W., & Khan, M. U. (2024). Nanomaterials in the treatment of degenerative intellectual and developmental disabilities. *Exploration of BioMat-X*, 1(6), 353-365. <https://doi.org/10.37349/ebmx.2024.00024>

Awadelkareem, A. M., Siddiqui, A. J., Noumi, E., Ashraf, S. A., Hadi, S., Snoussi, M., Badraoui, R., Bardakci, F., Ashraf, M. S., Danciu, C., Patel, M., & Adnan, M. (2023). Biosynthesized Silver Nanoparticles Derived from Probiotic *Lactobacillus rhamnosus* (AgNPs-LR) Targeting Biofilm Formation and Quorum Sensing-Mediated Virulence Factors. *Antibiotics* (Basel), 12(6), 986. <https://doi.org/10.3390/antibiotics12060986>

Buffet, P. E., Zalouk-Vergnoux, A., Chatel, A., Berthet, B., Mettais, I., Perrein-Ettajani, H., Poirier, L., Luna-Acosta, A., Thomas-Guyon, H., Risso-de Faverney, C., Guibbolini, M., Gilliland, D., Valsami-Jones, E., & Mouneyrac, C. (2014). A marine mesocosm study on the environmental fate of silver nanoparticles and toxicity effects on two endobenthic species: the ragworm *Hediste diversicolor* and the bivalve mollusc *Scrobicularia plana*. *Sci Total Environ*, 470-471, 1151-1159. <https://doi.org/10.1016/j.scitotenv.2013.10.114>

Cao, J., Feng, Y., He, S., & Lin, X. (2017). Silver nanoparticles deteriorate the mutual interaction between maize (*Zea mays L.*) and arbuscular mycorrhizal fungi: a soil microcosm study. *Applied Soil Ecology*, 119, 307-316. <https://doi.org/10.1016/j.apsoil.2017.04.011>

Chaachouay, N., Zidane, L., & Husen, A. (2024). Impact of Silver Nanoparticles in Aquatic Plants. In *Plant Response to Silver Nanoparticles: Plant Growth, Development, Production, and Protection* (pp. 249-263). Springer. [https://doi.org/10.1007/978-981-97-7352-7\\_14](https://doi.org/10.1007/978-981-97-7352-7_14)

Courtois, P. (2020). Ecotoxicological assessment of silver nanoparticles and their derivatives: their effects on fauna, flora and soil microorganisms [Université de Lille].

Dang, F., Huang, Y., Wang, Y., Zhou, D., & Xing, B. (2021). Transfer and toxicity of silver nanoparticles in the food chain. *Environmental Science: Nano*, 8(6), 1519-1535. <https://doi.org/10.1039/D0EN01190H10.1039/D0EN01190H>

De Matteis, V., Malvindi, M. A., Galeone, A., Brunetti, V., De Luca, E., Kote, S., Kshirsagar, P., Sabella, S., Bardi, G., & Pompa, P. P. (2015). Negligible particle-specific toxicity mechanism of silver nanoparticles: the role of Ag<sup>+</sup> ion release in the cytosol. *Nanomedicine*, 11(3), 731-739. <https://doi.org/10.1016/j.nano.2014.11.00210.1016/j.nano.2014.11.002>

de Oca-Vásquez, G. M., Solano-Campos, F., Vega-Baudrit, J. R., López-Mondéjar, R., Odriozola, I., Vera, A., Moreno, J. L., & Bastida, F. (2020). Environmentally relevant concentrations of silver nanoparticles diminish soil microbial biomass but do not alter enzyme activities or microbial diversity. *Journal of Hazardous Materials*, 391, 122224. <https://doi.org/10.1016/j.jhazmat.2020.122224>

Dodds, W. K., Guinnip, J. P., Schechner, A. E., Pfaff, P. J., & Smith, E. B. (2021). Fate and toxicity of engineered nanomaterials in the environment: A meta-analysis. *Science of the Total Environment*, 796, 148843. <https://doi.org/10.1016/j.scitotenv.2021.148843>

Du, J., Tang, J., Xu, S., Ge, J., Dong, Y., Li, H., & Jin, M. (2018). A review on silver nanoparticles-induced ecotoxicity and the underlying toxicity mechanisms. *Regulatory Toxicology and Pharmacology*, 98, 231-239. <https://doi.org/10.1016/j.yrtph.2018.08.003>

Eivazi, F., & Afraziabi, Z. (2018). Effects of silver nanoparticles on the activities of soil enzymes involved in carbon and nutrient cycling. *Pedosphere*, 28(2), 209-214. [https://doi.org/10.1016/S1002-0160\(18\)60019-0](https://doi.org/10.1016/S1002-0160(18)60019-0)

Ellis, L. A., Valsami-Jones, E., Lead, J. R., & Baalousha, M. (2016). Impact of surface coating and environmental conditions on the fate and transport of silver nanoparticles in the aquatic environment. *Sci Total Environ*, 568, 95-106. <https://doi.org/10.1016/j.scitotenv.2016.05.199>

Flores-Lopez, L. Z., Espinoza-Gomez, H., & Somanathan, R. (2019). Silver nanoparticles: Electron transfer, reactive oxygen species, oxidative stress, beneficial and toxicological effects. *Mini review. J Appl Toxicol*, 39(1), 16-26. <https://doi.org/10.1002/jat.3654> 10.1002/jat.3654

Furtado, L. M., Norman, B. C., Xenopoulos, M. A., Frost, P. C., Metcalfe, C. D., & Hintemann, H. (2015). Environmental Fate of Silver Nanoparticles in Boreal Lake Ecosystems. *Environ Sci Technol*, 49(14), 8441-8450. <https://doi.org/10.1021/acs.est.5b01116>

Gavin, A. (2016). Investigating the mechanisms of silver nanoparticle toxicity in *Daphnia magna*: a multi-omics approach, University of Birmingham.

Ghobashy, M. M., Abd Elkodous, M., Shabaka, S. H., Younis, S. A., Alshangiti, D. M., Madani, M., Al-Gahtany, S. A., Elkhateib, W. F., Noreddin, A. M., & Nady, N. (2021). An

overview of methods for production and detection of silver nanoparticles, with emphasis on their fate and toxicological effects on human, soil, and aquatic environment. *Nanotechnology Reviews*, 10(1), 954-977. <https://doi.org/10.1515/ntrev-2021-0066> 10.1515/ntrev-2021-0066

Gomez-Gomez, B., Arregui, L., Serrano, S., Santos, A., Perez-Corona, T., & Madrid, Y. (2019). Unravelling mechanisms of bacterial quorum sensing disruption by metal-based nanoparticles. *Sci Total Environ*, 696, 133869. <https://doi.org/10.1016/j.scitotenv.2019.133869>

Grün, A.-L., Manz, W., Kohl, Y. L., Meier, F., Straskraba, S., Jost, C., Drexel, R., & Emmerling, C. (2019). Impact of silver nanoparticles (AgNP) on soil microbial community depending on functionalization, concentration, exposure time, and soil texture. *Environmental Sciences Europe*, 31(1), 1-22. <https://doi.org/10.1186/s12302-019-0196-y>

Grün, A. L., Scheid, P., Hauröder, B., Emmerling, C., & Manz, W. (2017). Assessment of the effect of silver nanoparticles on the relevant soil protozoan genus Acanthamoeba. *Journal of Plant Nutrition and Soil Science*, 180(5), 602-613. <https://doi.org/10.1002/jpln.201700277>

He, D. (2013). Biotic and abiotic interactions of silver nanoparticles: aggregation, dissolution and reactive oxygen species generation [UNSW Sydney].

He, S., Feng, Y., Ni, J., Sun, Y., Xue, L., Feng, Y., Yu, Y., Lin, X., & Yang, L. (2016). Different responses of soil microbial metabolic activity to silver and iron oxide nanoparticles. *Chemosphere*, 147, 195-202. <https://doi.org/10.1016/j.chemosphere.2015.12.055>

Jahan, S., Yusoff, I. B., Alias, Y. B., & Bakar, A. F. B. A. (2017). Reviews of the toxicity behavior of five potential engineered nanomaterials (ENMs) into the aquatic ecosystem. *Toxicology Reports*, 4, 211-220. <https://doi.org/10.1016/j.toxrep.2017.04.001>

Jangid, H., Singh, S., Kashyap, P., Singh, A., & Kumar, G. (2024). Advancing biomedical applications: an in-depth analysis of silver nanoparticles in antimicrobial, anticancer, and wound healing roles. *Front Pharmacol*, 15, 1438227. <https://doi.org/10.3389/fphar.2024.1438227>

Kalantzi, I., Mylona, K., Toncelli, C., Bucheli, T. D., Knauer, K., Pergantis, S. A., Pitta, P., Tsiala, A., & Tsapakis, M. (2019). Ecotoxicity of silver nanoparticles on plankton organisms: a review. *Journal of nanoparticle research*, 21, 1-26. <https://doi.org/10.1007/s11051-019-4504-7>

Kang, J., Zhou, N., Zhang, Y.-w., Wang, Y.-h., Song, C.-q., Gao, X., Song, G.-f., Guo, J.-s., Huang, L., & Ma, T.-f. (2023). Synthesis, multi-site transformation fate and biological toxicity of silver nanoparticles in aquatic environment: A review. *Environmental Technology & Innovation*, 32, 103295. <https://doi.org/10.1016/j.eti.2023.103295>

Khan, M. R., & Akram, M. (2020). Nanoparticles and their fate in soil ecosystem. Biogenic nano-particles and their use in agro-ecosystems, 221-245. [https://doi.org/10.1007/978-981-15-2985-6\\_13](https://doi.org/10.1007/978-981-15-2985-6_13)

Khan, M. U., Ullah, H., Honey, S., Talib, Z., Abbas, M., Umar, A., Ahmad, T., Sohail, J., Sohail, A., & Makgopa, K. (2023). Metal nanoparticles: Synthesis approach, types and applications—a mini review. *Nano-Horizons: Journal of Nanosciences and Nanotechnologies*, 2, 21 pages-21 pages. <https://doi.org/10.1007/s00210-024-03082-y>

Khusro, A., Aarti, C., & Arasu, M. V. (2023). Biosurfactants-mediated Nanoparticles as Next-Generation Therapeutics. In *Multifunctional Microbial Biosurfactants* (pp. 455-494). Springer. [https://doi.org/10.1007/978-3-031-31230-4\\_21](https://doi.org/10.1007/978-3-031-31230-4_21)

Kulikova, N. (2021). Silver nanoparticles in soil: input, transformation, and toxicity. *Eurasian Soil Science*, 54, 352-365. <https://doi.org/10.1134/S1064229321030091>

Kurwadkar, S., Pugh, K., Gupta, A., & Ingole, S. (2015). Nanoparticles in the environment: Occurrence, distribution, and risks. *Journal of Hazardous, Toxic, and Radioactive Waste*, 19(3), 04014039. [https://doi.org/10.1061/\(ASCE\)HZ.2153-5515.0000258](https://doi.org/10.1061/(ASCE)HZ.2153-5515.0000258)

Kusi, J., & Maier, K. J. (2022). Evaluation of silver nanoparticle acute and chronic effects on freshwater amphipod (*Hyalella azteca*). *Aquat Toxicol*, 242, 106016. <https://doi.org/10.1016/j.aquatox.2021.106016>

Kwok, K. W., Auffan, M., Badireddy, A. R., Nelson, C. M., Wiesner, M. R., Chilkoti, A., Liu, J., Marinakos, S. M., & Hinton, D. E. (2012). Uptake of silver nanoparticles and toxicity to early life stages of Japanese medaka (*Oryzias latipes*): effect of coating materials. *Aquat Toxicol*, 120-121, 59-66. <https://doi.org/10.1016/j.aquatox.2012.04.012>

Lapresta-Fernández, A., Fernández, A., & Blasco, J. (2012). Nanoecotoxicity effects of engineered silver and gold nanoparticles in aquatic organisms. *TrAC Trends in Analytical Chemistry*, 32, 40-59. <https://doi.org/10.1016/j.trac.2011.09.007>

León-Silva, S., Fernández-Luqueño, F., & López-Valdez, F. (2016). Silver nanoparticles (AgNP) in the environment: a review of potential risks on human and environmental health. *Water, Air, & Soil Pollution*, 227(9), 306. <https://doi.org/10.1007/s11270-016-3022-9>

Li, C., Liu, Z., Xu, Y., Chen, X., Zhang, Q., Hu, L., Lv, Z., Liu, X., Xiao, T., & Li, D. (2024). AgNPs-induced oxidative stress and inflammation confer an increased susceptibility to aquatic reovirus infection. *Aquaculture*, 586, 740748. <https://doi.org/10.1016/j.aquaculture.2024.740748>

Li, P., Su, M., Wang, X., Zou, X., Sun, X., Shi, J., & Zhang, H. (2020). Environmental fate and behavior of silver nanoparticles in natural estuarine systems. *J Environ Sci (China)*, 88, 248-259. <https://doi.org/10.1016/j.jes.2019.09.013>

Li, Y., Qin, T., Ingle, T., Yan, J., He, W., Yin, J. J., & Chen, T. (2017). Differential genotoxicity mechanisms of silver nanoparticles and silver ions. *Arch Toxicol*, 91(1), 509-519. <https://doi.org/10.1007/s00204-016-1730-y>

McGee, C. F. (2020). The effects of silver nanoparticles on the microbial nitrogen cycle: a review of the known risks. *Environ Sci Pollut Res Int*, 27(25), 31061-31073. <https://doi.org/10.1007/s11356-020-09548-9>

Noga, M., Milan, J., Frydrych, A., & Jurowski, K. (2023). Toxicological Aspects, Safety Assessment, and Green Toxicology of Silver Nanoparticles (AgNPs)-Critical Review: State of the Art. *Int J Mol Sci*, 24(6), 5133. <https://doi.org/10.3390/ijms24065133>

Oktarina, H. (2017). The effect of silver nanoparticles on *Trichoderma harzianum*, *Rhizoctonia* spp., and fungal soil communities [Newcastle University].

Ottoni, C. A., Lima Neto, M. C., Leo, P., Ortolan, B. D., Barbieri, E., & De Souza, A. O. (2020). Environmental impact of biogenic silver nanoparticles in soil and aquatic organisms. *Chemosphere*, 239, 124698. <https://doi.org/10.1016/j.chemosphere.2019.124698>

Paciorek, P., Źuberek, M., & Grzelak, A. (2020). Products of lipid peroxidation as a factor in the toxic effect of silver nanoparticles. *Materials*, 13(11), 2460. <https://doi.org/10.3390/ma13112460>

Padhye, L. P., Jasemizad, T., Bolan, S., Tsyusko, O. V., Unrine, J. M., Biswal, B. K., Balasubramanian, R., Zhang, Y., Zhang, T., Zhao, J., Li, Y., Rinklebe, J., Wang, H., Siddique, K. H. M., & Bolan, N. (2023). Silver contamination and its toxicity and risk management in terrestrial and aquatic ecosystems. *Sci Total Environ*,

871, 161926.  
<https://doi.org/10.1016/j.scitotenv.2023.161926>

Peixoto, S., Loureiro, S., & Henriques, I. (2022). The impact of silver sulfide nanoparticles and silver ions in soil microbiome. *J Hazard Mater*, 422, 126793. <https://doi.org/10.1016/j.jhazmat.2021.126793>

Peyrot, C., Wilkinson, K. J., Desrosiers, M., & Sauvé, S. (2014). Effects of silver nanoparticles on soil enzyme activities with and without added organic matter. *Environmental Toxicology and Chemistry*, 33(1), 115-125. <https://doi.org/10.1002/etc.2398> 10.1002/etc.2398

Qadeer, B., Khan, M. A., Tariq, H., Zahid, M. U., Alismail, H. A. A., Hussain, S. J., Ahmad, U., & Bokhari, S. A. I. (2024). PEGylation of silver nanoparticles via Berginia Ciliata aqueous extract for biological applications. *Emergent Materials*, 7(4), 1657-1673. <https://doi.org/10.1007/s42247-024-00727-9>

Qi, L., Li, Z., Liu, J., & Chen, X. (2024). Omics-Enhanced Nanomedicine for Cancer Therapy. *Adv Mater*, 36(50), e2409102. <https://doi.org/10.1002/adma.202409102>

Rajkuberan, C., Sudha, K., Sathishkumar, G., & Sivaramakrishnan, S. (2015). Antibacterial and cytotoxic potential of silver nanoparticles synthesized using latex of *Calotropis gigantea* L. *Spectrochim Acta A Mol Biomol Spectrosc*, 136 Pt B, 924-930. <https://doi.org/10.1016/j.saa.2014.09.115>

Rajput, V., Minkina, T., Ahmed, B., Sushkova, S., Singh, R., Soldatov, M., Laratte, B., Fedorenko, A., Mandzhieva, S., Blicharska, E., Musarrat, J., Saquib, Q., Flieger, J., & Gorovtsov, A. (2020). Interaction of Copper-Based Nanoparticles to Soil, Terrestrial, and Aquatic Systems: Critical Review of the State of the Science and Future Perspectives. *Rev Environ Contam Toxicol*, 252, 51-96. [https://doi.org/10.1007/398\\_2019\\_34](https://doi.org/10.1007/398_2019_34)

Saini, P., Saha, S. K., Roy, P., Chowdhury, P., & Babu, S. P. S. (2016). Evidence of reactive oxygen species (ROS) mediated apoptosis in *Setaria cervi* induced by green silver nanoparticles from *Acacia auriculiformis* at a very low dose. *Experimental parasitology*, 160, 39-48. <https://doi.org/10.1016/j.exppara.2015.11.004>

Shah, S., Gaikwad, S., Nagar, S., Kulshrestha, S., Vaidya, V., Nawani, N., & Pawar, S. (2019). Biofilm inhibition and anti-quorum sensing activity of phytosynthesized silver nanoparticles against the nosocomial pathogen *Pseudomonas aeruginosa*. *Biofouling*, 35(1), 34-49. <https://doi.org/10.1080/08927014.2018.1563686>

Shaikh, W. A., Chakraborty, S., Owens, G., & Islam, R. U. (2021). A review of the phytochemical mediated synthesis of AgNP (silver nanoparticle): the wonder particle of the past decade. *Appl Nanosci*, 11(11), 2625-2660. <https://doi.org/10.1007/s13204-021-02135-5>

Sharma, V. K., Sayes, C. M., Guo, B., Pillai, S., Parsons, J. G., Wang, C., Yan, B., & Ma, X. (2019). Interactions between silver nanoparticles and other metal nanoparticles under environmentally relevant conditions: A review. *Science of the Total Environment*, 653, 1042-1051. <https://doi.org/10.1016/j.scitotenv.2018.10.411>

Singh, K., Thakur, S. S., Ahmed, N., Alharby, H. F., Al-Ghamdi, A. J., Al-Solami, H. M., Bahattab, O., & Yadav, S. (2022). Ecotoxicity assessment for environmental risk and consideration for assessing the impact of silver nanoparticles on soil earthworms. *Heliyon*, 8(10), e11167. <https://doi.org/10.1016/j.heliyon.2022.e11167>

Sohn, E. K., Johari, S. A., Kim, T. G., Kim, J. K., Kim, E., Lee, J. H., Chung, Y. S., & Yu, I. J. (2015). Aquatic Toxicity Comparison of Silver Nanoparticles and Silver Nanowires. *Biomed Res Int*, 2015(1), 893049. <https://doi.org/10.1155/2015/893049>

Tangaa, S. R., Selck, H., Winther-Nielsen, M., & Khan, F. R. (2016). Trophic transfer of metal-based nanoparticles in aquatic environments: a review and recommendations for future research focus. *Environmental Science: Nano*, 3(5), 966-981. <https://doi.org/10.1039/C5EN00280J>

Tonczyk, A., Niedzialkowska, K., & Lisowska, K. (2025). Ecotoxic effect of mycogenic silver nanoparticles in water and soil environment. *Sci Rep*, 15(1), 10815. <https://doi.org/10.1038/s41598-025-95485-x>

Tortella, G., Rubilar, O., Durán, N., Diez, M., Martínez, M., Parada, J., & Seabra, A. (2020). Silver nanoparticles: Toxicity in model organisms as an overview of its hazard for human health and the environment. *Journal of Hazardous Materials*, 390, 121974. <https://doi.org/10.1016/j.jhazmat.2019.121974>

Wang, P., Lombi, E., Menzies, N. W., Zhao, F.-J., & Kopittke, P. M. (2018). Engineered silver nanoparticles in terrestrial environments: a meta-analysis shows that the overall environmental risk is small. *Environmental Science: Nano*, 5(11), 2531-2544. <https://doi.org/10.1039/C8EN00486B>

Wolska, K. I., Grudniak, A. M., & Markowska, K. (2017). Inhibition of bacterial quorum sensing systems by metal nanoparticles. *Metal Nanoparticles in Pharma*, 123-138. [https://doi.org/10.1007/978-3-319-63790-7\\_7](https://doi.org/10.1007/978-3-319-63790-7_7)

Xiao, H., Chen, Y., & Alnaggar, M. (2019). Silver nanoparticles induce cell death of colon cancer cells through impairing cytoskeleton and membrane nanostructure. *Micron*, 126, 102750. <https://doi.org/10.1016/j.micron.2019.102750>

Yadav, A., & Sahu, D. (2024). Synthesis of Silver Nanoparticles from Plant Extracts and Their Potential Applications in Cancer Treatment: A Comprehensive Review. *International Journal of Pharmacognosy and Herbal Drug Technology (IJPHDT)*, 1(1), 53-76.

Yang, Y.-F., Cheng, Y.-H., & Liao, C.-M. (2017). Nematode-based biomarkers as critical risk indicators on assessing the impact of silver nanoparticles on soil **ecosystems**. *Ecological Indicators*, 75, 340-351. <https://doi.org/10.5555/20173107586>

Zhang, H., Huang, M., Zhang, W., Gardea-Torresdey, J. L., White, J. C., Ji, R., & Zhao, L. (2020). Silver Nanoparticles Alter Soil Microbial Community Compositions and Metabolite Profiles in Unplanted and Cucumber-Planted Soils. *Environ Sci Technol*, 54(6), 3334-3342. <https://doi.org/10.1021/acs.est.9b07562>

Zhang, W., Ke, S., Sun, C., Xu, X., Chen, J., & Yao, L. (2019). Fate and toxicity of silver nanoparticles in freshwater from laboratory to realistic environments: a review. *Environ Sci Pollut Res Int*, 26(8), 7390-7404. <https://doi.org/10.1007/s11356-019-04150-0>