# ASSOCIATION OF CLASS I HUMAN LEUKOCYTE ANTIGEN IN IRAQI PATIENTS WITH HODGKIN'S LYMPHOMA

RUQAYA MUHAMMED AL-BARZINJI Hawler Medical University,College of Medicine, Kurdistan Region-Iraq (Accepted for publication: June 9, 2013)

#### ABSTRACT

#### **Objective:-**

To assess the correlation between human leukocytes antigen class I in Iraqi patients with Hodgkin's lymphoma. <u>Subject and Methods:-</u>

A study was conducted in period between the June 2003 and November, 2004. Study groups included 80 newly diagnosed Hodgkin's lymphoma patients and two control groups, patient controls which include 50 newly diagnosed by Non Hodgkin's lymphoma and 50 healthy controls. Antibody-Mediated complement dependent cytotoxicity assay was done by treating sample of patient's lymphocytes with a panel of anti- human leukocyte antigen antisera and complement.

Results:-

The frequency expression of human leukocyte antigen- A1, A28, B5, B51 and Cw7 was significantly greater in Hodgkin's lymphoma patients compared to healthy control group with (p < 0.001) for human leukocyte antigen- A1, A28 and Cw7 and (p < 0.01) for B5 and B51.

#### Conclusion:-

Human leukocyte antigen- A1, A28, B5, B51 and Cw7 are more related with Hodgkin's lymphoma in Iraqi population that reflects allelic association to Hodgkin's lymphoma.

Key word: Class I human leukocyte antigen , Hodgkin's lymphoma

### **INTRODUCTION**

Hodgkin's lymphoma (HL) is a form of malignant lymphoma, that is unusual in that the bulk of the tumor is composed of normal cells within which the malignant Reed-Sternberg (RS cells) lie (Derek *et al.*, 1995). Such classic binucleated (owls eyes) malignant cells in all histologic subtypes usually express antigens found on resting or activated lymphocytes (Urba *et al.*, 1992).

While the specific cause of HL is unknown, clusters of the disease in certain regions have been noted, and both genetic and infectious processes are suspected (Ambindar, 2002 and Jarrett, 2003).

In Iraq, among the common malignancies HL ranks the tenth (Ministry of Health, 1993, 1996 and 1999), while in Gulf countries and south east governorate of Yemen, HL is the fourth common cancer (Rabadi , 1987; Ezzata *et al.*, 1996 and Bawazir, 1998).

The human leukocyte antigen classes (HLA) region has been subdivided into class I, class II, and class III regions. Each region contains numerous gene loci, and each locus may encode a large number of polymorphic alleles. Class I and class II antigens are composed of two polypeptide chains, associated together by non-

covalent link, usually called alpha and beta chains, which dimerize to form the final molecule (Hyde, 2000).

Strong arguments supporting a genetic linkage between susceptibility to HL and HLA antigens (Hors *et al.*,1983) . Moreover, other studies denote that HL is determined by both an HLAassociated major gene and other non-HLA genetic factors together with environmental effects (Shugart *et al.*, 2000).

# PATIENTS AND METHODS

### Patients: -

Cross sectional study was conducted in the following study groups in period between the Jun 2003, and November, 2004. The patients included in this case - control study were classified into 2 groups, patients and control groups.

Eighty newly diagnosed HL patients (32 females and 48 males) who were either attending the Institute of Radiology and Nuclear Medicine or admitted to AL-Mansour and Baghdad Teaching Hospitals in Baghdad City.

All patients were in their new onset of the disease (not on chemo or radiotherapy) at which the histopathological samples were taken and diagnosed as HL according to the National Cancer Institute Working Formula. All patients were subjected to personal interview using especially designed questionnaire format.

Control groups were age, sex and ethnic matched with patients group, they consisted of two groups patient control which included 50 patients who were newly diagnosed to be affected by NHL and healthy control which included 50 healthy individuals whom were not complaining of any malignant problem.

# **METHODS**

Ten ml of venous blood were drawn from each subject (patients, patients controls and healthy controls). The venous blood was dispensed into plastic or glass universal tubes containing either lithium heparin (10 Iu/ml blood) as anti- coagulant, or glass beads followed by a gentile mixing for HLA typing.

Typing of HLA -A, B and C antigens was carried out in the tissue typing laboratory, Al -Karama hospital, and in the Teaching Laboratories of Medical City in Baghdad .The test microlymphocytoxicity was established by (Terasaki and MaClelland, 1964) and modified by (Dick *et al.*, 1979 and Bender, 1984).

Antibody-Mediated complement dependent cytotoxicity assay was done by treating sample of patient's lymphocytes with a panel of anti-HLA antisera and complement.

Anti- HLA sera react with the corresponding lymphocyte antigens without visible cell alteration .The addition of rabbit complement, leads to a change in the structure of lymphocyte cell membrane which can be made visible by means of an indicator vital dye (eosin). The lysed and vital lymphocytes were assessed using an inverse phase contrast microscope. The significance of an association between HLA alleles and both patients and control calculated using the Chi-square test with Yates correction as well as Fisher exact test (Dorak *et al.*, ,2002). To determine the strength of association between HLA specificities and disease, the relative risk (RR), etiological fraction (EF), preventive fraction (PF) and type of association were estimated.

### RESULTS

In this study, the HLA class I (A, B and C) allele frequency were determined by microlymphocytotoxicity assay in the 2 groups of lymphoma patients. The first group included 80 patients with HL, while the second included 50 individuals with NHL considered as patients control. Their allele frequencies were compared with 50 healthy controls. The HLA frequencies were compared by Fisher exact test.

Human leukocyte antigen (HLA) typing results obtained for HLA-Class I antigens are summarized in tables (1, 2, 3, 4, 5 and 6). The frequencies of HLA- A1, A28, B5, B51 and Cw7 were significantly increased in HL patients (37.5, 21.2, 37.5, 28.7 and 43.7% respectively) compared with healthy controls (p < 0.001) for HLA- A1, A28 and Cw7 and (p < 0.01) for B5 and B51 (Tables 1, 2 and 3). Such positive associations were presented with RR values of 5, 3.7, 4, 5.4 and 6.4, respectively and EF values of 0.299, 0.155, 0.244, 0.234 and, 0.369, respectively with positive association.

In comparison between results of HL patients with NHL patients control as shown in tables (4, 5 and 6), it was found that the HLA-Cw6 frequency is significantly (p<0.04) increased in NHL patient control than in HL patient (44% versus 28%) ( p <0.05).

HLA –A antigens	cor	Healthy control No. 50		control		control		control		control		oatients o. 80	RR	Ρ	EF	PF	Type of Association
	Ν	%	Ν	%													
1	5	10.0	30	37.5	5	0.0003	0.299		PA								
2	24	48.0	22	27.5	0.4	NS		0.278	NA								
3	14	28.0	15	18.7	0.6	NS		0.112	NA								
9	7	14.0	9	11.2	0.7	NS		0.032	NA								
10	4	8.0	11	13.7	1.7	NS	0.057		PA								
11	3	6.0	5	6.2	0.9	NS		0.0007	NA								
23	3	6.0	4	5.0	0.7	NS		0.012	NA								
24	3	6.0	4	5.0	0.7	NS		0.012	NA								

 Table 1: Frequency of HLA-A antigens in Hodgkin lymphoma patients and healthy control

25	1	2.0	2	2.5	1.0	NS	0.001		PA
26	2	4.0	1	1.2	0.3	NS		0.021	NA
28	3	6.0	17	21.2	3.7	0.0008	0.155		PA
29	2	4.0	3	3.7	0.8	NS		0.005	NA
30	5	10.0	3	3.7	0.3	NS		0.059	NA
31	1	2.0	4	5.0	1.9	NS	0.024		PA
32	1	2.0	2	2.5	1.0	NS	0.001		PA
33	5	10.0	2	2.5	0.2	NS		0.065	NA
34	3	6.0	6	7.5	1.1	NS	0.011		PA
36	3	6.0	3	3.7	0.6	NS		0.023	NA
Blank	11		17						
Tetal	100		16						
Total	100		0						

RR = Relative Risk; EF= Etiologic Fraction; PF=Preventive fraction ; PA=Positive Association; (EF> 0.15); NA=Negative Association(PF>0.15); NS=Non Significant; -- = Nill ; P=Probability

**Table 2 :** Frequency of HLA-B antigens in Hodgkin lymphoma patients and healthy control

HLA –B antigens	Hea con No	trol		patients lo 80	RR	Ρ	EF	PF	Type of Associati
	No.	%	No.	%					on
5	5	10.0	26	37.5	4.0	0.0024	0.244		PA
7	2	4.0	8	10.0	2.2	NS	0.056		PA
8	4	8.0	14	17.5	2.2	NS	0.097		PA
12	3	6.0	1	1.2	0.2	NS		0.035	NA
13	1	2.0	2	2.5	1.0	NS	0.001		PA
14	3	6.0	1	1.2	0.2	NS		0.035	NA
15	1	2.0	2	2.5	1.0	NS	0.001		PA
16	3	6.0	4	5.0	0.7	NS		0.012	NA
17	2	4.0	2	2.5	0.6	NS		0.015	NA
18	3	6.0	1	1.2	0.2	NS		0.035	NA
21	5	10.0	4	5.0	0.4	NS		0.050	NA
22	1	2.0	5	6.2	2.4	NS	0.036		PA
27	1	2.0	1	1.2	0.6	NS		0.007	NA
35	5	10.0	4	0.0	0.4	NS		0.050	NA
37	0	0.0	0	0.0	0.6	NS	0.0	0.0	ND *
38	2	4.0	2	2.5	0.6	NS		0.015	NA
39	2	4.0	1	1.2	0.3	NS		0.021	NA
40	3	6.0	1	1.2	0.2	NS		0.035	NA
41	3	6.0	1	1.2	0.2	NS		0.035	NA
44	4	8.0	2	2.5	0.3	NS		0.048	NA
45	2	4.0	3	3.7	0.8	NS		0.005	NA
47	1	2.0	3	3.7	1.4	NS	0.012		PA
48	0	0.0	1	1.2	1.9	NS	0.005		PA
49	2	4.0	4	5.0	1.1	NS	0.006		PA
50	2	4.0	1	1.2	0.3	NS		0.021	NA
51	4	8.0	23	28.7	5.4	0.0032	0.234		PA
52	1	2.0	3	3.7	1.4	NS	0.012		PA
53	1	2.0	0	0.0	0.2	NS	0.0	0.0	ND
54	1	2.0	2	2.5	1.0	NS	0.001		PA
55	2	4.0	0	0.0	0.1	NS	0.0	0.0	ND

2 2 2	4.0 4.0 4.0	2 2	2.5 2.5	0.6 0.6	NS NS		0.015	NA		
	-	_	2.5	0.6	NS		0.015	NLA		
2	10	0			110		0.015	NA		
	4.0	2	2.5	0.6	NS		0.015	NA		
1	2.0	1	1.2	0.6	NS		0.007	NA		
1	2.0	2	2.5	1.0	NS	0.001		PA		
2	4.0	3	3.7	0.8	NS		0.005	NA		
1	2.0	5	6.2	2.4	NS	0.036		PA		
20		21								
100	* ND: Non determined									
	1 20	1     2.0       2     4.0       1     2.0       20     20	1     2.0     2       2     4.0     3       1     2.0     5       20     21	1         2.0         2         2.5           2         4.0         3         3.7           1         2.0         5         6.2           20         21         21	1       2.0       2       2.5       1.0         2       4.0       3       3.7       0.8         1       2.0       5       6.2       2.4         20       21       21       21       21	1         2.0         2         2.5         1.0         NS           2         4.0         3         3.7         0.8         NS           1         2.0         5         6.2         2.4         NS           20         21         21         20         21         20	1         2.0         2         2.5         1.0         NS         0.001           2         4.0         3         3.7         0.8         NS            1         2.0         5         6.2         2.4         NS         0.036           20         21         21         21         21         21         21         21	1       2.0       2       2.5       1.0       NS       0.001          2       4.0       3       3.7       0.8       NS        0.005         1       2.0       5       6.2       2.4       NS       0.036          20       21       21       21       21       21       21       20       21		

Table 3: Frequency of HLA-C antigens in Hodgkin lymphoma patients and healthy control

HLA –C antigens	Hea con No.	,	lymphoma patients		RR	Ρ	EF	PF	Type of associa tion
	No.	%	No.	%					
1	4	8.0	5	6.3	0.7	NS		0.020	NA
2	8	16.0	9	11.3	0.6	NS		0.053	NA
3	7	14.0	6	7.5	0.5	NS		0.068	NA
4	13	26.0	16	20.0	0.7	NS		0.075	NA
5	2	4.0	3	3.8	0.8	NS		0.005	NA
6	11	22.0	22	28.0	1.3	NS	0.066		PA
7	5	10.0	35	43.7	6.4	0.00002	0.369		PA
8	1	2.0	3	3.8	1.4	NS	0.012		PA
Blank	49		61						
Total	100		160						

Table 4: Frequency of HLA-A antigens in Hodgkin lymphoma patients and Non-Hodgkin lymphoma control

HLA –A antigens		control . 50		patients lo. 80	Р
	Ν	%	N	%	
1	23	46	30	37.5	NS
2	15	30	22	27.5	NS
3	11	22	15	18.7	NS
9	4	8	9	11.2	NS
10	8	16	11	13.7	NS
11	2	4	5	6.2	NS
23	1	2	4	5.0	NS
24	3	6	4	5.0	NS
25	2	4	2	2.5	NS
26	3	6	1	1.2	NS
28	9	18	17	21.2	NS
29	1	2	3	3.7	NS
30	6	12	3	3.7	NS
31	1	2	4	5.0	NS
32	2	4	2	2.5	NS
33	3	6	2	2.5	NS
34	2	4	6	7.5	NS
36	1	2	3	3.7	NS
Blank	3		17		
Total	100		160		

HLA –B antigens		IL control No. 50	Н	L patients No. 80	Ρ	
	No.	%	No.	%	-	
5	19	38.0	26	37.5	NS	
7	9	18.0	8	10.0	NS	
8	6	12.0	14	17.5	NS	
12	1	2.0	1	1.2	NS	
13	1	2.0	2	2.5	NS	
14	0	0.0	1	1.2	NS	
15	1	2.0	2	2.5	NS	
16	2	4.0	4	5.0	NS	
17	1	2.0	2	2.5	NS	
18	0	0.0	1	1.2	NS	
21	1	2.0	4	5.0	NS	
22	1	2.0	5	6.2	NS	
27	0	0.0	1	1.2	NS	
35	2	4.0	4	0.0	NS	
37	1	2.0	0	0.0	NS	
38	1	2.0	2	2.5	NS	
39	0	0.0	1	1.2	NS	
40	0	0.0	1	1.2	NS	
41	2	4.0	1	1.2	NS	
44	1	2.0	2	2.5	NS	
45	1	2.0	3	3.7	NS	
47	1	2.0	3	3.7	NS	
48	0	0.0	1	1.2	NS	
49	3	6.0	4	5.0	NS	
50	0	0.0	1	1.2	NS	
51	15	30.0	23	28.7	NS	
52	1	2.0	3	3.7	NS	
53	1	2.0	0	0.0	NS	
54	1	2.0	2	2.5	NS	
55	1	2.0	0	0.0	NS	
56	1	2.0	2	2.5	NS	
57	2	4.0	2	2.5	NS	
60	1	2.0	2	2.5	NS	
62	0	0.0	1	1.2	NS	
63	2	4.0	2	2.5	NS	
70	1	2.0	3	3.7	NS	
73	2	4.0	5	6.2	NS	
Blank	18		21			
Total	100		160			

 Table 5: Frequency of HLA-B antigens in Hodgkin lymphoma patients and non-Hodgkin lymphoma control

HLA –C antigens	NH	HL control No.50	Н	Р	
	Ν	%	Ν	%	
1	2	4.0	5	6.3	NS
2	8	16.0	9	11.3	NS
3	4	8.0	6	7.5	NS
4	12	24.0	16	20.0	NS
5	1	2.0	3	3.8	NS
6	22	44.0	22	28.0	0.041
7	17	34.0	35	43.7	NS
8	3	6.0	3	3.8	NS
Blank	31		61		
Total	100		160		

**Table 6:** Frequency of HLA-C antigens in Hodgkin lymphoma patients and non- Hodgkin lymphoma control

### DISCUSSION

Hodgkin's lymphoma is relatively rare lymphoma that affects younger as well as elder persons (Mueller, 1996). The disease aggregation in families and persons with specific HLA types indicates genetic susceptibility (Glaser *et al.*, 1996).

Human Leukocyte Antigen is located on the short arm of chromosome 6, due to its extreme polymorphism it is considered as one of the best genetic markers and the most voluble prognostic factors used nowadays (Fogdell *et al.*, 1998).

A number of reports have studied the associations between HL and HLA, some of them established correlation between several antigens and HL while other found no correlations. The present study is the first to determine the frequencies of HLA-class I using lymphocytotoxicity test in Iraqi patients with HL. Examination has been done for 80 HL patients, 50 NHL patients control and 50 healthy controls. The present study revealed that in patients with HL there were significantly increase in frequency for certain alleles, A1, A28, B5, B51 and Cw7 in HLA-class I, as compared to healthy control, which may confirm the existence of antigenic linkage between such alleles and the susceptibility to the HL in certain individuals.

The frequencies of class I HLA-A1, 28, B5, 51 and Cw7 were increased significantly in HL patients compared to healthy control, however, non of these alleles showed significant positive or negative association with NHL patient control excluded Cw6 allele that showed significant negative association in NHL patients compared to patients with HL.

Different from our results, Amiel, (1967); Green et al., (1979) and Hors et al., (1983) found significant differences in the HLA- B8 allele while they found B5 allele were found to be similar to that of our study. Racial differences and methods of analysis may be the key factors for the controversy of these results from ours. In addition, 60% to 70% of cases in familial HL may be linked to this region (Chakravorti et al., 1986). In 1993, Conte et al., in their work on familial cases of NS subtype of HL, found that B18 allele is related to the development of familial HL, particularly the familial NS subtype, however the incidence of HLA-B locus antigens especially that involved B5, B15, B18 and B35 was more associated with HL patients(Tiwari et al., 1980). Xixiong et al., (1996) reported that HLA28, B18, B51, Cw4 and DQ4 alleles significantly increased as compared NHL patients with healthy control. In addition, the association of HL with different alleles as evident in this work and other published studies may be explained by more than one possibility. One implies that genes determining these conditions are located close to the MHC loci, and are thus transmitted along with whatever HLA haplotypes in the family (Hafez et al., 1985). On the other word heritable

factors contribute to development of HL (Jox et

al., 2002).A second possibility is the cross-

tolerance to a self competent (Woda et al.,

1981). A third possibility is that the immunogenic responsiveness to oncogenic viruses may be linked to genes coding for HLA antigens, if the concerned coding of the key antigens is missing or present and active, the person will become susceptible to the virus. In conclusion, The susceptibility to HL in Iraqi population is supposed to be related to certain specific alleles(HLA- A1, A28, B5, B51 and Cw7) which suggests that certain HLA alleles may be associated with predisposition to HD.

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پەيوەندى نێوان پۆلى يەكەمى ئەنتيجينى ليوكوسايتى مرۆڤى بە نەخۆشە عيراقىيە تووش بووەكان بە نەخۆشى ھۆچكنى ليمفاوى

پوخته

ئامانج: بۆ زانينى پەيوەندى نێوان پۆلى يەكەمى ئەنتىجينى ليوكوسايتى مرۆڤى بە نەخۆشە عيراقىيە تووش بورەكان بە نەخۆشى ھۆچكنى ليمفاوى ھەيە

A1, A28, ئەنجامى ئەم لىكۆلىينەوەيە دەرىخىىت كەوا دووبارەبوونەوەى ئەنتىجينى ليوكوسايتى مرۆڤى پۆلى ,A28 A1, A28 B5, B51,Cw7 لە نەخۆشى ھۆچكنى ليمفاوى زياترە بە بەراوورد كردن لەگەل ھەردوو كۆنترۆلەكە و جياوازي ديارو بەلگەدارى ئاماركارى تۆماركرا بە رادەى (p <0.001) لەگەل پۆلى ,A1, A28, Cw7 . وە بە رادەى (p <0.01) لە لەگەل پۆلى B5, B51 دەردانى ئەم دوو ماددەيە لەلايەن ميزەللدان رۆلىكى گرنگ دەبينن لە نەخۆشى ھۆچكنى ليمفاوى

دەرئەنجام: ئەنتىجىنى ليوكوسايتى مرۆڤى پۆلى A1, A28, B5, B51,Cw7 زۆرتر پەيوەندىدار بوون بە نەخۆشى ھۆچكنى ليمفاوى لە خەلكانى عيراق، ئەمەش رۆلى ئەليلى ئەبەستىنت بە نەخۆشىيەكەوە. ووشە سەرەكيەكان: ئەنتىجىنى ليوكوسايتى مرۆڤى پۆلى يەكەم، لە نەخۆشى ھۆچكنى ليمفاوى Cancer Institute Working Formula. All patients were subjected to personal interview using especially designed questionnaire format.

Control groups were age, sex and ethnic matched with patients group, they consisted of two groups patient control which included 50 patients who were newly diagnosed to be affected by NHL and healthy control which included 50 healthy individuals whom were not complaining of any malignant problem.

# **METHODS**

Ten ml of venous blood were drawn from each subject (patients, patients controls and healthy controls). The venous blood was dispensed into plastic or glass universal tubes containing either lithium heparin (10 Iu/ml blood) as anti- coagulant, or glass beads followed by a gentile mixing for HLA typing.

Typing of HLA -A, B and C antigens was carried out in the tissue typing laboratory, Al -Karama hospital, and in the Teaching Laboratories of Medical City in Baghdad .The test microlymphocytoxicity was established by (Terasaki and MaClelland, 1964) and modified by (Dick *et al.*, 1979 and Bender, 1984).

Antibody-Mediated complement dependent cytotoxicity assay was done by treating sample of patient's lymphocytes with a panel of anti-HLA antisera and complement.

Anti- HLA sera react with the corresponding lymphocyte antigens without visible cell alteration .The addition of rabbit complement, leads to a change in the structure of lymphocyte cell membrane which can be made visible by means of an indicator vital dye (eosin). The lysed and vital lymphocytes were assessed using an inverse phase contrast microscope. The significance of an association between HLA alleles and both patients and control calculated using the Chi-square test with Yates correction as well as Fisher exact test (Dorak *et al.*, ,2002). To determine the strength of association between HLA specificities and disease, the relative risk (RR), etiological fraction (EF), preventive fraction (PF) and type of association were estimated.

## RESULTS

In this study, the HLA class I (A, B and C) allele frequency were determined by microlymphocytotoxicity assay in the 2 groups of lymphoma patients. The first group included 80 patients with HL, while the second included 50 individuals with NHL considered as patients control. Their allele frequencies were compared with 50 healthy controls. The HLA frequencies were compared by Fisher exact test.

Human leukocyte antigen (HLA) typing results obtained for HLA-Class I antigens are summarized in tables (1, 2, 3, 4, 5 and 6). The frequencies of HLA- A1, A28, B5, B51 and Cw7 were significantly increased in HL patients (37.5, 21.2, 37.5, 28.7 and 43.7% respectively) compared with healthy controls (p < 0.001) for HLA- A1, A28 and Cw7 and (p < 0.01) for B5 and B51 (Tables 1, 2 and 3). Such positive associations were presented with RR values of 5, 3.7, 4, 5.4 and 6.4, respectively and EF values of 0.299, 0.155, 0.244, 0.234 and, 0.369, respectively with positive association.

In comparison between results of HL patients with NHL patients control as shown in tables (4, 5 and 6), it was found that the HLA-Cw6 frequency is significantly (p<0.04) increased in NHL patient control than in HL patient (44% versus 28%) ( p <0.05).

HLA –A antigens	cor	althy htrol . 50		patients p. 80	RR	Р	EF	PF	Type of Association
	Ν	%	Ν	%					
1	5	10.0	30	37.5	5	0.0003	0.299		PA
2	24	48.0	22	27.5	0.4	NS		0.278	NA
3	14	28.0	15	18.7	0.6	NS		0.112	NA
9	7	14.0	9	11.2	0.7	NS		0.032	NA
10	4	8.0	11	13.7	1.7	NS	0.057		PA
11	3	6.0	5	6.2	0.9	NS		0.0007	NA
23	3	6.0	4	5.0	0.7	NS		0.012	NA
24	3	6.0	4	5.0	0.7	NS		0.012	NA

 Table 1: Frequency of HLA-A antigens in Hodgkin lymphoma patients and healthy control

25	1	2.0	2	2.5	1.0	NS	0.001		PA
26	2	4.0	1	1.2	0.3	NS		0.021	NA
28	3	6.0	17	21.2	3.7	0.0008	0.155		PA
29	2	4.0	3	3.7	0.8	NS		0.005	NA
30	5	10.0	3	3.7	0.3	NS		0.059	NA
31	1	2.0	4	5.0	1.9	NS	0.024		PA
32	1	2.0	2	2.5	1.0	NS	0.001		PA
33	5	10.0	2	2.5	0.2	NS		0.065	NA
34	3	6.0	6	7.5	1.1	NS	0.011		PA
36	3	6.0	3	3.7	0.6	NS		0.023	NA
Blank	11		17						
Total	100		16						
iolai	100		0						

RR = Relative Risk; EF= Etiologic Fraction; PF=Preventive fraction ; PA=Positive Association; (EF> 0.15); NA=Negative Association(PF>0.15); NS=Non Significant; -- = Nill ; P=Probability

**Table 2 :** Frequency of HLA-B antigens in Hodgkin lymphoma patients and healthy control

HLA –B antigens	antigens No 5			patients lo 80	RR	Ρ	EF	PF	Type of Associati
	No.	%	No.	%					on
5	5	10.0	26	37.5	4.0	0.0024	0.244		PA
7	2	4.0	8	10.0	2.2	NS	0.056		PA
8	4	8.0	14	17.5	2.2	NS	0.097		PA
12	3	6.0	1	1.2	0.2	NS		0.035	NA
13	1	2.0	2	2.5	1.0	NS	0.001		PA
14	3	6.0	1	1.2	0.2	NS		0.035	NA
15	1	2.0	2	2.5	1.0	NS	0.001		PA
16	3	6.0	4	5.0	0.7	NS		0.012	NA
17	2	4.0	2	2.5	0.6	NS		0.015	NA
18	3	6.0	1	1.2	0.2	NS		0.035	NA
21	5	10.0	4	5.0	0.4	NS		0.050	NA
22	1	2.0	5	6.2	2.4	NS	0.036		PA
27	1	2.0	1	1.2	0.6	NS		0.007	NA
35	5	10.0	4	0.0	0.4	NS		0.050	NA
37	0	0.0	0	0.0	0.6	NS	0.0	0.0	ND *
38	2	4.0	2	2.5	0.6	NS		0.015	NA
39	2	4.0	1	1.2	0.3	NS		0.021	NA
40	3	6.0	1	1.2	0.2	NS		0.035	NA
41	3	6.0	1	1.2	0.2	NS		0.035	NA
44	4	8.0	2	2.5	0.3	NS		0.048	NA
45	2	4.0	3	3.7	0.8	NS		0.005	NA
47	1	2.0	3	3.7	1.4	NS	0.012		PA
48	0	0.0	1	1.2	1.9	NS	0.005		PA
49	2	4.0	4	5.0	1.1	NS	0.006		PA
50	2	4.0	1	1.2	0.3	NS		0.021	NA
51	4	8.0	23	28.7	5.4	0.0032	0.234		PA
52	1	2.0	3	3.7	1.4	NS	0.012		PA
53	1	2.0	0	0.0	0.2	NS	0.0	0.0	ND
54	1	2.0	2	2.5	1.0	NS	0.001		PA
55	2	4.0	0	0.0	0.1	NS	0.0	0.0	ND

56	2	4.0	2	2.5	0.6	NS		0.015	NA	
57	2	4.0	2	2.5	0.6	NS		0.015	NA	
60	2	4.0	2	2.5	0.6	NS		0.015	NA	
62	1	2.0	1	1.2	0.6	NS		0.007	NA	
63	1	2.0	2	2.5	1.0	NS	0.001		PA	
70	2	4.0	3	3.7	0.8	NS		0.005	NA	
73	1	2.0	5	6.2	2.4	NS	0.036		PA	
Blank	20		21							
Total	100	160 * ND: Non determined								

Table 3: Frequency of HLA-C antigens in Hodgkin lymphoma patients and healthy control

HLA –C antigens	con	Healthy control No. 50		Hodgkin lymphoma patients No. 80		Ρ	EF	PF	Type of associa tion
	No.	%	No.	%					
1	4	8.0	5	6.3	0.7	NS		0.020	NA
2	8	16.0	9	11.3	0.6	NS		0.053	NA
3	7	14.0	6	7.5	0.5	NS		0.068	NA
4	13	26.0	16	20.0	0.7	NS		0.075	NA
5	2	4.0	3	3.8	0.8	NS		0.005	NA
6	11	22.0	22	28.0	1.3	NS	0.066		PA
7	5	10.0	35	43.7	6.4	0.00002	0.369		PA
8	1	2.0	3	3.8	1.4	NS	0.012		PA
Blank	49		61						
Total	100		160						

Table 4: Frequency of HLA-A antigens in Hodgkin lymphoma patients and Non-Hodgkin lymphoma control

HLA –A antigens		control o. 50	HL N	Р	
	N	%	N	%	
1	23	46	30	37.5	NS
2	15	30	22	27.5	NS
3	11	22	15	18.7	NS
9	4	8	9	11.2	NS
10	8	16	11	13.7	NS
11	2	4	5	6.2	NS
23	1	2	4	5.0	NS
24	3	6	4	5.0	NS
25	2	4	2	2.5	NS
26	3	6	1	1.2	NS
28	9	18	17	21.2	NS
29	1	2	3	3.7	NS
30	6	12	3	3.7	NS
31	1	2	4	5.0	NS
32	2	4	2	2.5	NS
33	3	6	2	2.5	NS
34	2	4	6	7.5	NS
36	1	2	3	3.7	NS
Blank	3		17		
Total	100		160		

HLA –B antigens	NHL control No. 50		Н	Р	
	No.	%	No.	%	-
5	19	38.0	26	37.5	NS
7	9	18.0	8	10.0	NS
8	6	12.0	14	17.5	NS
12	1	2.0	1	1.2	NS
13	1	2.0	2	2.5	NS
14	0	0.0	1	1.2	NS
15	1	2.0	2	2.5	NS
16	2	4.0	4	5.0	NS
17	1	2.0	2	2.5	NS
18	0	0.0	1	1.2	NS
21	1	2.0	4	5.0	NS
22	1	2.0	5	6.2	NS
27	0	0.0	1	1.2	NS
35	2	4.0	4	0.0	NS
37	1	2.0	0	0.0	NS
38	1	2.0	2	2.5	NS
39	0	0.0	1	1.2	NS
40	0	0.0	1	1.2	NS
41	2	4.0	1	1.2	NS
44	1	2.0	2	2.5	NS
45	1	2.0	3	3.7	NS
47	1	2.0	3	3.7	NS
48	0	0.0	1	1.2	NS
49	3	6.0	4	5.0	NS
50	0	0.0	1	1.2	NS
51	15	30.0	23	28.7	NS
52	1	2.0	3	3.7	NS
53	1	2.0	0	0.0	NS
54	1	2.0	2	2.5	NS
55	1	2.0	0	0.0	NS
56	1	2.0	2	2.5	NS
57	2	4.0	2	2.5	NS
60	1	2.0	2	2.5	NS
62	0	0.0	1	1.2	NS
63	2	4.0	2	2.5	NS
70	1	2.0	3	3.7	NS
73	2	4.0	5	6.2	NS
Blank	18		21		
Total	100		160		

Table 5: Frequency of HLA-B antigens in Hodgkin lymphoma patients and non-Hodgkin lymphoma control

HLA –C antigens	NHL control No.50		Н	Р	
	Ν	%	Ν	%	
1	2	4.0	5	6.3	NS
2	8	16.0	9	11.3	NS
3	4	8.0	6	7.5	NS
4	12	24.0	16	20.0	NS
5	1	2.0	3	3.8	NS
6	22	44.0	22	28.0	0.041
7	17	34.0	35	43.7	NS
8	3	6.0	3	3.8	NS
Blank	31		61		
Total	100		160		

**Table 6:** Frequency of HLA-C antigens in Hodgkin lymphoma patients and non- Hodgkin lymphoma control

### DISCUSSION

Hodgkin's lymphoma is relatively rare lymphoma that affects younger as well as elder persons (Mueller, 1996). The disease aggregation in families and persons with specific HLA types indicates genetic susceptibility (Glaser *et al.*, 1996).

Human Leukocyte Antigen is located on the short arm of chromosome 6, due to its extreme polymorphism it is considered as one of the best genetic markers and the most voluble prognostic factors used nowadays (Fogdell *et al.*, 1998).

A number of reports have studied the associations between HL and HLA, some of them established correlation between several antigens and HL while other found no correlations. The present study is the first to determine the frequencies of HLA-class I using lymphocytotoxicity test in Iraqi patients with HL. Examination has been done for 80 HL patients, 50 NHL patients control and 50 healthy controls. The present study revealed that in patients with HL there were significantly increase in frequency for certain alleles, A1, A28, B5, B51 and Cw7 in HLA-class I, as compared to healthy control, which may confirm the existence of antigenic linkage between such alleles and the susceptibility to the HL in certain individuals.

The frequencies of class I HLA-A1, 28, B5, 51 and Cw7 were increased significantly in HL patients compared to healthy control, however, non of these alleles showed significant positive or negative association with NHL patient control excluded Cw6 allele that showed significant negative association in NHL patients compared to patients with HL.

Different from our results, Amiel, (1967); Green et al., (1979) and Hors et al., (1983) found significant differences in the HLA- B8 allele while they found B5 allele were found to be similar to that of our study. Racial differences and methods of analysis may be the key factors for the controversy of these results from ours. In addition. 60% to 70% of cases in familial HL may be linked to this region (Chakravorti et al., 1986). In 1993, Conte et al., in their work on familial cases of NS subtype of HL, found that B18 allele is related to the development of familial HL, particularly the familial NS subtype, however the incidence of HLA-B locus antigens especially that involved B5, B15, B18 and B35 was more associated with HL patients(Tiwari et al., 1980). Xixiong et al., (1996) reported that HLA28, B18, B51, Cw4 and DQ4 alleles significantly increased as compared NHL patients with healthy control. In addition, the association of HL with different alleles as evident in this work and other published studies may be explained by more than one possibility. One implies that genes determining these conditions are located close to the MHC loci, and are thus transmitted along with whatever HLA haplotypes in the family (Hafez et al., 1985). On the other word heritable factors contribute to development of HL (Jox et

al., 2002). A second possibility is the cross-

tolerance to a self competent (Woda et al.,

1981). A third possibility is that the immunogenic responsiveness to oncogenic viruses may be linked to genes coding for HLA antigens, if the concerned coding of the key antigens is missing or present and active, the person will become susceptible to the virus. In conclusion, The susceptibility to HL in Iraqi population is supposed to be related to certain specific alleles(HLA- A1, A28, B5, B51 and Cw7) which suggests that certain HLA alleles may be associated with predisposition to HD.

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پەيوەندى نێوان پۆلى يەكەمى ئەنتيجينى ليوكوسايتى مرۆڤى بە نەخۆشە عيراقىيە تووش بووەكان بە نەخۆشى ھۆچكنى ليمفاوى

پوخته

ئامانج: بۆ زانينى پەيوەندى نێوان پۆلى يەكەمى ئەنتيجينى ليوكوسايتى مرۆڤى بە نەخۆشە عيراقىيە تووش بورەكان بە نەخۆشى ھۆچكنى ليمفاوى ھەيە

A1, A28, ئەنجامى ئەم ليكۆلىينەوەيە دەرىخىىت كەوا دووبارەبوونەوەى ئەنتيجينى ليوكوسايتى مرۆڤى پۆلى ,A28 A1, A28 م B5, B51,Cw7 لە نەخۆشى ھۆچكنى ليمفاوى زياترە بە بەراوورد كردن لەگەل ھەردوو كۆنترۆلەكە و جياوازي ديارو بەلگەدارى ئاماركارى تۆماركرا بە رادەى (p <0.001) لەگەل پۆلى ,A1, A28, Cw7 وہ به رادەى (p <0.01) لە لەگەل پۆلى B5, B51 دەردانى ئەم دوو ماددەيە لەلايەن ميزەلدان رۆليكى گرنگ دەبينن لە نەخۆشى ھۆچكنى ليمفاوى

دەرئەنجام: ئەنتىجىنى ليوكوسايتى مرۆڤى پۆلى A1, A28, B5, B51,Cw7 زۆرتر پەيوەندىدار بوون بە نەخۆشى ھۆچكنى ليمفاوى لە خەلكانى عيراق، ئەمەش رۆلى ئەليلى ئەبەستىنت بە نەخۆشىيەكەوە. ووشە سەرەكيەكان: ئەنتىجىنى ليوكوسايتى مرۆڤى پۆلى يەكەم، لە نەخۆشى ھۆچكنى ليمفاوى