

MOLECULAR IDENTIFICATION AND HEMOLYTIC ACTIVITY OF CANDIDA SPECIES ISOLATED FROM URINE OF HEALTHY AND DIABETIC WOMEN IN KURDISTAN OF IRAQ

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ABSTRACT:

A survey was carried out on the incidence of Candida yeasts in urine of healthy (non-diabetic) and diabetic women in Duhok and Zakho cities from February 2019 to April 2019. Out of 620 urine samples examined, 144 samples were found positive for Candida infection (23.2%). There were significant differences in incidence of Candida infection between urine of healthy and diabetic women as well as among age groups. All Candida isolates from positive samples were identified by using Chromogenic Candida Agar medium and confirmed by sequencing of the ITS1 and ITS4 region of rDNA. The identified Candida species were *C.albicans*, *C.glabrata*, *C.krusei* and *C.tropicalis*. The most common species was *C.albicans* (34%), followed by *C.glabrata* (29.9%), then *C.tropicalis* (13.8%), whereas, *C.krusei* (= *Pichia kudriavzevii*) displayed the least incidence (9.7%). All tested strains (n=83) for their hemolytic activity on Sabouraud's dextrose agar amended with sheep blood and glucose showed positive activity. *Candida albicans* and *C.tropicalis* displayed the highest hemolytic activity.

KEYWORDS: Hemolysin, Candida spp., Urine, Diabetic Mellitus, Sequencing, ITS.

1. INTRODUCTION

One of the most common mycobiota in human which colonizing several functional sites like skin, mouth, urinary tract, vagina is *Candida* without causing any infection (Seneviratne et al., 2008). Modification in local environment of the host enhanced *Candida* growth and hence led to candidiasis (Pope and Cole, 2002). Virulence factors of *Candida* as well as immune system of the host switching *Candida* from harmless commensally to disease causing pathogen (Yang, 2003). Most common virulence factors of *Candida*; adherence and extracellular hydrolase enzymes which help the *Candida* to adapt to specific anatomical sites. The pathogenesis of *Candida* species enhanced by phenotypic switching, filamentation and biofilm formation (Calderone and Fonzi, 2001). Extracellular enzymes like as proteases, phospholipases, haemolysin and lipase contributed in pathogenicity of *Candida* species (Silva et al., 2009).

Furthermore, the capability of *C. albicans* to get iron by haemolysin production is essential in its persistence and capability to form infections in the human's body. Hemolytic activity is one of the virulence factors that facilitate hyphal invasion and disseminate candidiasis. The activity of hemolysin by *Candida* is enhanced by present of glucose in the blood agar (Manns et al., 1994). Hemolysins production by *Candida albicans* destroy hemoglobin in erythrocytes and obtain elemental iron. Hence, hemolysins considered as essential virulence factors which make pathogens to survive and persist (Luo et al., 2004).

The most suitable medium for study the hemolytic activity of *Candida* isolates is Sabouraud's dextrose agar with addition of Sheep blood. The role of CaCl₂ on the hemolysin activity of *Candida* was studied by Linares and his colleagues (Linares et al., 2007). The addition of 2.5 % CaCl₂ to Sabouraud's

dextrose agar contains sheep blood reduce the hemolytic activity of *C.dubliniensis* while the *C. albicans* strains were stimulated (Koga-Ito et al., 2006). Unicellular and multicellular organisms need essential elements for growth such as iron, which present in the hemoglobin within red blood cells in the human body. *C.albicans* binding to complement receptors of erythrocytes by mannoprotein on the cell surface of yeast then produces hemolysin which lysis of the erythrocyte (Moors et al., 1992; Watanabe et al., 1999 and Almeida et al., 2009).

Various studies showed significant relationship between high incidence of *Candida* in urine samples and predisposing factors such as pregnancy state of woman, Diabetic mellitus, prolong used of antibiotics and used bladder catheters (Khudor et al., 2002; Falahati et al., 2016 and Lima et al., 2017). Several studies for diagnosis of *Candida* species from urine of diabetic and non-diabetic women revealed that there is significant relationship between high level of glucose in blood (diabetic patients) and high prevalence of Candiduria (Pandey and Pandey, 2013; and Yismaw et al., 2013). Although, several diagnostic methods such as phenotypic methods (germ tube, Chlamydo spores production and Chromogenic Candida Agar) were used, molecular analysis technique such as Multiplex PCR and Sequenced PCR are seem to be more accurate and precise methods for identification of *Candida* species from urine and vaginal samples (Mohammed and Said, 2015; Lima et al., 2017). Intergenic Spacer Region (ITS) is placed among the constant genes coding for 28S rRNA and 18S rRNA, while 5.8SrRNA gene is highly conserved and located between ITS1 and ITS4 (White et al., 1990). The genetic diversity of ITS1 and ITS4 regions give well identification of closely related species of *Candida* (Ciardo et al., 2006). The aim of the present study was isolation and identification of *Candida* species from urine of diabetic and non-diabetic women by using molecular

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analysis techniques and study the hemolytic activity of some *Candida* isolates.

2. MATERIALS AND METHODS

2.1. Samples Collection

Urine samples (n= 620) were collected from healthy and diabetic women attended laboratory of Maternity hospital and Diabetes Center in Zakho and Duhok city. The study was achieved between February 2019 and April 2019. Ten ml of urine samples were kept in sterilized universal containers. Patient information was noted such as age, types of therapy (antibiotic and antifungal) and other possible risk factors. All urine samples were transported to mycology laboratory at Zakho University for analysis.

2.2. Isolation and phenotypic Identification of *Candida* Species

Samples were cultured within few hours on Sabouraud's dextrose agar (SDA) amended with 0.05% (Weight/Volume) chloramphenicol for inhibiting bacteria growth by using streak plate method (Bhavan et al., 2010). Cultures incubated at 37°C for two days after that pure *Candida* colonies were sub-cultured on Harlequin TM *Candida* Chromogenic agar (CHROM agar Company, Acumedia, Neogen, UK) at 35°C for 48 hours. Presumptive identification of *Candida* strains was done depend on color of the colonies (Pfaller et al., 1996).

2.3. Determination of Hemolysin activity

Eighty three isolates of *Candida* recovered from urine specimens were checked for hemolytic activity according to Manns et al. (Manns et al., 1994). To determine the hemolytic activity; Sabouraud's dextrose agar enriched with 7 ml of sheep blood was inoculated with 10⁸ cells/ml suspension of each isolated *Candida* species and incubated for one day at 37°C. To estimate the effect of glucose on the production of hemolysis, 10 µl of each sample's suspension inoculated on two SDA medium containing blood; one enriched with 3% glucose and other without glucose within two days at 37 ° C in the presence of 5% CO₂ atmosphere. The activity of Hemolysis by different *Candida* species was determined by measure hemolytic index (Hz value) as seen in the following equation:

$$Hz = \frac{\text{translucent zone of haemolysis (mm)}}{\text{Colony diameter (mm)}}$$

A reference strain of *Staphylococcus aureus* was used as a positive control for beta hemolytic activity (Luo et al., 2001).

2.4. Molecular Analysis

2.4.1. DNA Extraction: A loop-full from each *Candida* isolate was taken and cultured on disposable sterile tube (2 ml) containing 1 ml Yeast Extract Peptone Dextrose (YPD) Broth which had been prepared previously and then incubated in shaking incubator at 37°C for 24 hours. The genomic DNA extract using a method based on spin column genomic DNA purification using yeast DNA preparation kits (Jena Bioscience- Germany). All extracted genomic DNA was suspended in 50 µl of Elution Buffer (EB) and freeze at 4 °C until used. Purity and concentration of DNA, was measured by the Nanodrop.

2.4.2. PCR Amplification Technique: The non-specific fungal primers ITS1 (5'- TCCGTAGGTGAACCTGCGG-3') and ITS4 (5'-TCCTCCGCTTATTGATAT GC-3') were used to amplify the Internal Transcribed Spacer (ITS) of ribosomal DNA region (rDNA) (20). The PCR was performed in 30 µl reaction mixture consisting of approximately 15 µl of master mix, 2 µl of template DNA, 1 µl of forward primer ITS1 and 1 µl of reverse primers and 11 µl of PCR-grade water all of these

kept in a single tube. The PCR cycle parameters were as follows: an initial denaturation step was 95°C for 5 min. followed by 10 cycles of denaturation at 94°C for 30 seconds, annealing at 60°C for 45seconds, and extension at 72°C for 90 seconds and for 25 cyclers as followed; denaturation at 94°C for 30 seconds, annealing at 55°C for 45seconds, and extension at 72°C for 90seconds then final extension at 72°C for 10 minutes. PCR products were analyzed by agarose gel electrophoresis in 1X TBE buffer at 100 V for 120 min in gel composed of 1.5% agarose. 100bp ladder DNA Marker was run with PCR products for sizing of the bands. Gels were stained with Red Safe DNA loading dye, then visualized with a UV Transilluminator and photographed.

2.4.3. Sequencing of DNA: PCR products of fifteen *Candida* species isolates from the samples were sequenced using primers (ITS1 and ITS4) by Macrogen Company (Seoul, Korea). Basic Local Alignment Search Tool (BLAST) for sequence analysis databases was used to identify the obtained DNA sequences ([http:// www.ncbi.nlm.nih.gov/BLAST](http://www.ncbi.nlm.nih.gov/BLAST)). All sequences were clean up and align by BioEdit application.

2.5. Data Analysis

The data were analyzed using the SPSS software (Statistical Package for the Social Sciences, version (Kauffman et al., 2000). The relative proportions were calculated with a confidence interval of 95%. To determine the association between variables, the chi-squared (x²) test was used; a p – value < 0.05 was considered significant and more than that considered not significant.

3. RESULTS AND DISCUSSION

A total of 620 urine samples were analyzed for presence of *Candida* yeast, 144 samples showed positive results. In diabetic patients, the highest percentage was recorded for the age group 15-25 years (35.7%), followed by (32.0%) for age group 26-36 and (25%) for age group 48-60 years, whereas 12% was recorded for age group 37-47. The least ratio (4.5%) was recorded for the age group above 60 years. For non-diabetic women, the highest incidence of *Candida* infection (18%) was recorded for the age group 26-36 years, followed by (14.2%) for the age group 15-25 years and (4%) for the age group 37-47 year; p=0.01. We observed that there was significant association between age groups and the positive percentage of diabetes and non-diabetes women (Table 1).

The detection of *Candida* species in urine of diabetic and non-diabetic women could be contamination or reflection of colonization or infection of the urinary tract by *Candida* yeast (Kauffman et al., 2000 and Mohammed et al., 2017). In general, the incidence of *Candida* yeast isolated from urine of patients with diabetes were higher than non-diabetes patient and this can be explained due to the high level of glucose in urine of diabetic patients as well as the suppression of the immune system in diabetic women (Falahati et al., 2016). The highest incidence of *Candida* yeasts (35.7% and 32%) was found in the age groups of 15-25 years and 26-36 years in diabetic patients respectively. Higher incidence (14.2% and 18%) was also reported for the age groups 18-25 years and 26-36 years respectively. This finding was in agreement with other studies (Navin et al., 2004; Geerlings et al., 2014; Falahati et al., 2016).

Table 1. Incidence percentage of Candida in urine samples from diabetic and healthy women

| Age | Number of samples examined | % of positive samples in diabetes women | % of positive samples in non-diabetes women | % of Positive samples |
|--|----------------------------|---|---|-----------------------|
| 15-25 | 70 | 25/70 (35.7 %) | 10/70 (14.2 %) | 35/70 (50 %) |
| 26-36 | 100 | 32/100 (32 %) | 18/100 (18 %) | 50/100 (50 %) |
| 37 - 47 | 150 | 18/150 (12 %) | 6/150 (4 %) | 24/150 (16 %) |
| 48-60 | 100 | 25/100 (25 %) | 0 (0%) | 25/100 (25 %) |
| 60 < | 200 | 9/200 (4.5%) | 1/200 (0.5%) | 10/200 (5 %) |
| Total | 620 (100 %) | 109/620 (17.5 %) | 35/620 (5.6 %) | 144/620 (23.2 %) |
| P value = 0.010 analyzed using the SPSS software using the chi-squared (x2) test | | | | |

All positive Candida isolates were cultured (n=144) on chromogenic Candida agar for presumptive identification of Candida species. Chromogenic Candida agar medium detected 131 isolated out of 144; including 49 isolates of *C. albican* (green), 43 isolates of *C. glabrata* (white shiny), and 14 isolates of *C. krusei* (purple to pink) while 20 isolates were *C. tropicalis* (blue). Mix infection with *C. albicans* and *C. glabrata* were observed in urine of our diabetes patients while only one non diabetes patient harbored two Candida species, the p value was 0.096 which is more than p value = 0.05 hence, we did not observe any significant associations among different species of Candida variables with Diabetes and Non diabetes and other disease (Table2).

The highest incidence displayed by *C.albicans* (34%) followed by *C.glabrata* (29.9%) and *C.tropicalis* (13.8%), whereas, the least incidence was recorded for *C.krusei* (9.7%), 3.5% of Mix infection with *C. albicans* and *C. glabrata*. Thirteen samples were not detected on chromogenic candida agar (Table 2).

This is in line with other different studies that reported a prevalence for *C.albicans*, *C.glabrata* and *C.tropicalis* (Achkar and Fries 2010; Ozhak-Baysan et al., 2012 and Ortiz et al.,2018). The three Candida species have been reported as pathogens in urinary tract infections (Papon et al., 2013). In contrast, *C.krusei* was found with high prevalence (26.31%) in urine samples collected from patients with urinary tract infection from Duhok (Mohammed et al., 2017). A mixed Candida infection was found in four urine samples from diabetic women and one sample from non-diabetic patient. Our result is in agreement with other studies that reported more than one Candida species (Kauffman et al.,2000; Mohammed et al., 2017 and Ortiz et al., 2018).

Table 2. Detection of Candida species among Diabetic and non-diabetic patients by using Chromogenic Candida Agar

| Species | NDM No (%) | DM No (%) | DM& other disease*No (%) | Total positive No (%) |
|----------------------|--------------|---------------|--------------------------|-----------------------|
| <i>C.albican</i> | 16/50 (32%) | 10/50 (20%) | 23/50 (46%) | 49/144 (34) |
| <i>C. glabrata</i> | 9/43 (20.9%) | 14/43 (32.6%) | 20/43 (46.5%) | 43/144 (29.9 %) |
| <i>C. tropicalis</i> | 5/20 (25%) | 5/20 (25%) | 10/20 (50%) | 20/144 (13.8%) |
| <i>C. krusei</i> | 4/14 (28.5%) | 6/14 (42.8%) | 4/14 (28.5%) | 14/144 (9.7%) |

| | | | | |
|---|----------------|----------------|----------------|-----------------|
| <i>C.albican</i> & <i>C.glabrata</i> | 1/4 (25%) | 4/4 (100%) | 0/4 (0%) | 5/144 (3.5%) |
| Not detected on chromo agar | 3/13 (23%) | 4/13 (30.7%) | 6/13 (46.1%) | 13/144 (9%) |
| <i>Candida</i> spp. identification | 35/131 (26.7%) | 39/131 (29.7%) | 57/131 (43.5%) | 131/144 (90.9%) |
| Total | 38/144 (26.4) | 43/144 (29.8) | 63/144 (43.8) | 144/144 (100%) |
| DM: Diabetes mellitus NDM: Non Diabetes mellitus *Other diseases = hypertension and cholesterol P value = 0.096 analyzed using the SPSS software using the chi-squared (x2) test | | | | |

All tested isolates of Candida have beta hemolytic activity, the hemolytic index of *Candida albicans* and *Candida tropicalis* were (1.80 ± 0.1) and (1.48 ± 0.159) respectively, which were higher than hemolytic index of *C. glabrata* (1.36 ± 0.129) and *C. krusei* (1.11 ± 0.101) as shown in Table3.

Table 3. Hemolytic Activity of Candida species on Blood Sheep Agar

| Species | No. of isolates with hemolysis (Hemolysis Index, mean ± SD) |
|---|---|
| | Beta |
| <i>Candida albicans</i> | 20 (1.80 ± 0.1) |
| <i>Candida glabrata</i> | 32 (1.36 ± 0.129) |
| <i>Candida tropicalis</i> | 18 (1.48 ± 0.159) |
| <i>Candida krusei</i> | 13 (1.11 ± 0.101) |
| All Data analyzed by using the SPSS software using the Standard Division SD P value = 0.091 | |

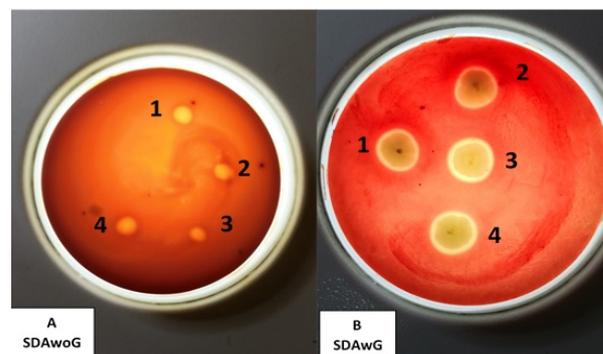


Figure 1. Hemolysis of Sheep blood agar induced by different species of Candida. A: Candida species on (SDAwG) Sabouraud's dextrose agar without glucose; B: Candida species on (SDAwG) Sabouraud's dextrose agar with glucose. 1. *C. albicans*; 2. *C. glabrata*; 3. *C. tropicalis*; 4. *C. krusei*.

Figure (1) showed the hemolytic activity of different species of Candida after 24 hours of inoculation. In the present study *C. albicans*, *C. glabrata*, *C. tropicalis* and *C. krusei* on Sabouraud's Dextrose blood supplemented with glucose (SDBwG) have high hemolytic activities compared to same strains cultured on Sabouraud Dextrose blood without glucose SDBwoG that showed negative hemolytic activity (Figure1).

One of the virulence factors of Candida is hemolysins. Species of Candida; *C. albicans* and *C. tropicalis* showed significantly beta-hemolytic activities higher than those of *C. glabrata*, and

C. krusei. In the current study, the hemolysis degree of all isolated *Candida* species was totally high in the medium containing glucose. The hemolytic activity of *Candida* was different regarding the present of glucose in the medium. *Candida albicans* is dimorphic microorganism which exist in two form hyphal and blastoconidial phases and that relay on the growth medium and condition. Some previous studies reported that *Candida* produced hemolysin only in the hyphal phases, our result do not agree with this finding as *C. glabrata* which is considered a hyphae-negative species exhibited both alpha and beta hemolysis (Paul et al., 1999).

Based on PCR results, universal primer ITS1 and ITS4 amplified DNA fragment from DNA template, PCR products size ranged from 500 to 1000 base pair. The results of DNA sequencing were aligned by BioEdit program and all edited sequences were submitted to Gene bank and compared with

other references strains from same host which have similarity ranged from 96% to 100% (Table 4). Data of DNA sequencing of *Candida* species were analyzed by Neighbor-Joining method using MEGA 7 (Tree View software) to construct phylogenetic and compared with references strain (Figure 2).

In the present study, specimens of *Candida* spp. were characterized by sequencing of the ITS-1 and ITS-4 rDNA, compared with references from different country such as china, Iran, Japan, Saudi Arabia, Kuwait and Egypt. The analysis revealed that the similarity ranged from 96% to 100%. The phylogenetic tree showed high relatedness among isolates of the same species of *Candiada*, they were generally assigned into two main clusters; most of them were belonged to the first cluster except *Candida glabrata* (MN50788) was clustered as monophyletic. In addition, this isolate has also a relationship with *Candida glabrata* (MN521700) with 76% bootstrap.

Table 4. List of internal transcribed spacers (ITS) sequences accession number and their similarities with our *Candida* isolates at Gen bank.

| Spp. | Accession number | Strain no. | Source | Location | Similarity |
|----------------------|------------------|------------|-------------------|--------------|------------|
| <i>C.tropicalis</i> | MN504645 | | Urine DM | IRAQ | 100 % |
| <i>C.tropicalis</i> | LT837794 | Kw3-15 | Urine | Kuwait | 100% |
| <i>C. tropicalis</i> | MN508369 | | Urine NDM | IRAQ | 100 % |
| <i>C.tropicalis</i> | KP674700 | h24b | Oral | China | 100% |
| <i>C.tropicalis</i> | KX015889 | AUMC 10251 | Gut | Egypt | 100% |
| <i>C. albicans</i> | MN519554 | 29 | Urine DM | IRAQ | 100 % |
| <i>C. albican</i> | MK793255 | Y94 | Clinical isolated | Iran | 99 % |
| <i>C. albican</i> | MK568486 | CA06.1 | Vaginal swab | Saudi Arabia | 99% |
| <i>C. albicans</i> | MN336231 | | Vulvovagina 1 | Iran | 100 % |
| <i>C. albican</i> | MN519597 | 44 | Urine NDM | IRAQ | 100 % |
| <i>C. albican</i> | MK564528 | CA-18G | Human/urine | Iran | 99% |
| <i>C. glabrata</i> | MN521700 | H35 | Urine DM | IRAQ | 98 % |
| <i>C. glabrata</i> | LC389242 | H7 | Vagina | Iran | 98% |
| <i>C. glabrata</i> | LC388879 | IFM 64903 | Clinical | Japan | 98% |
| <i>C. glabrata</i> | LC389246 | H35 | Vaginal | Iran | 99% |
| <i>C. glabrata</i> | MN507880 | | Urine NDM | IRAQ | 100% |
| <i>C. glabrata</i> | LC389256 | H108 | Vagina | Iran | 98% |
| <i>C. krusei</i> | MN515434 | m80b | Urine DM | IRAQ | 100 % |
| <i>C. krusei</i> | KP675284 | m80b | Oral | China | 97% |
| <i>C. krusei</i> | KP765013 | 121A | Gastric mucosa | China | 96% |
| <i>C. krusei</i> | KP675284 | m80b | Oral | China | 97 % |

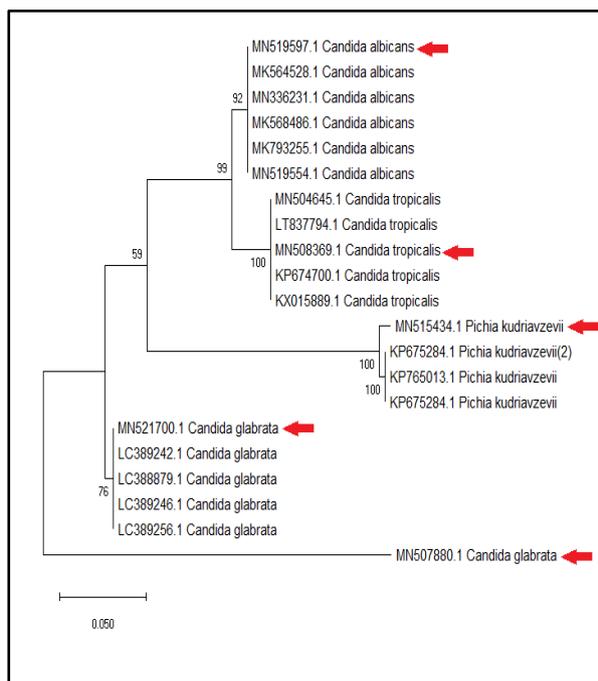


Figure 2. Phylogenetic tree of *Candida* spp. based on the ITS – rDNA sequencing.

Concerning the phylogenetic analysis for *Candida albicans* revealed that the obtained sequences shares 100% homology to *Candida albicans* strain: Iraqi isolate (MN519554), Iranian isolate (MN336231), 99% to both Iranian strains (MK793255 and MN564528) and Saudi Arabia isolate (MK568486). The phylogenetic analysis for *Candida tropicalis* showed that the obtained sequence shares 100% homology to *Candida tropicalis* strain: Iraqi isolate (MN504645), Kuwaiti isolate (LT837794), Chinese isolate (KP674700) and Egyptian isolate (KX015889). The morphological and molecular identification showed that the *Candida* isolates was *Candida tropicalis* with GeneBank accession number (MN508369).

The results from Table (4) and Figure (2) showed that the obtained sequence of *Pichia kudriavzevii* shares 97% homology to *Candida krusei* strain from China (KP675284 and PK675284) and 96% homology to the Chinese strain (KP765013). The morphological and molecular identification showed that the *Candida* isolates was *Candida krusei* with GeneBank accession number (MN515434). The phylogenetic tree analysis for the species *Candida glabrata* (GeneBank accession number: MN521700) showed that the obtained sequence shared 99% homology to *Candida glabrata* strain: Iranian isolate (LC389246), 98% homology to Japanese strain (LC388879) and Iranian strain (LC389242). While the isolate *Candida glabrata* (GeneBank accession number: MN507880) was clustered as monophyletic.

4. CONCLUSION

Candida albicans and non-*albicans* species were detected in urine of diabetic and non-diabetic women. All the isolated *Candida* species are important opportunistic pathogens. Our clinically relevant findings indicate that presence of glucose can increase the activity of hemolysins and all *Candida* species showed the beta hemolysis on blood agar and both *Candida albicans* and *Candida tropicalis* showed maximum hemolysis activity compared to *C. glabrata* and *C. krusei*.

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