

## THE EFFECT OF URTICA DIOICA EXTRACTS ON THIRD LARVAL INSTARS OF TRIBOLIUM CONFUSUM (DU VAL) USING TWO METHODS OF EXTRACTION

Mehvan M. Ali Rasheed<sup>a,\*</sup>, Badia'a Mahmoud Al-Chalabi<sup>a</sup>, Sofyan H. Sedo Al-Sinjari<sup>a</sup>

<sup>a</sup> Dept. of Biology, Faculty of Science, University of Zakho, Zakho, Kurdistan Region, Iraq – ([Mehvan.ali@stud.uoz.edu.krd](mailto:Mehvan.ali@stud.uoz.edu.krd); [badeyamahmoud50@gmail.com](mailto:badeyamahmoud50@gmail.com); [sofyan.sedo@uoz.edu.krd](mailto:sofyan.sedo@uoz.edu.krd))

Received: 17 Jul., 2022 / Accepted: 10 Oct., 2022 / Published: 07 Nov., 2022 <https://doi.org/10.25271/sjuoz.2022.10.4.974>

### ABSTRACT:

The effect of ethanolic extracts of *Urtica dioica* on mortality of the third Larval instar of *Tribolium confusum*, was studied using the Soxhlet and solvent extraction methods. Experiments at  $30 \pm 1^\circ\text{C}$  and  $70 \pm 5\%$  relative humidity. In laboratory conditions. The mortality data were recorded after 24, 48, and 72 hours. The mortality rate increased by an increase in concentrations (0.5, 1, 1.5, and 2.5 mg/ml), the highest mortality rate was 100% at 1.5 mg/ml, while the lowest percentage of mortality 73.33% was observed after 24 hrs of exposure at 0.5 mg/ml dose by Soxhlet extraction method, whereas the highest mortality rates 100% at 2.5 mg/ml concentration, while the lowest percentage of mortality 90% was observed after 24 hrs of exposure at 0.5 mg/ml dose by Solvent extraction methods. The results show that the plant extracts tested at different concentrations were toxic to the larvae of *T. confusum*, and both methods of extraction had nearly similar effects on larvae.

**KEYWORDS:** *Urtica dioica*, *Tribolium confusum*, Extraction methods, mortality rate.

### 1. INTRODUCTION

The confused flour beetle (CFB) (*Tribolium confusum* Du Val) of the family Tenebrionidae (order Coleoptera), is among the greatest common and harmful pests of stored products, such as flour mills, cereals, and other products worldwide (AL-Jamil and AL-Sinjari, 2019). Verheggen *et al.* (2007) state that their adults produce benzo-quinones and volatile hydrocarbons as defensive compounds that act as repellents on conspecific larvae and adults. Contamination of the human food supply is one of the most important dangers that people face (Al Abadi *et al.*, 2008). These pests can cause post-harvest losses of 9% or more in developed countries and 20% or more in developing countries (Phillips and Throne, 2010). Currently, methyl bromide and phosphine are the most often used fumigants for suppressing grain-infesting insects. However, their usage is restricted due to the detrimental impact they have on the environment, where the use of natural compounds as fumigants is an alternative technique for preventing insect infestations of stored products (Lee *et al.*, 2004). Botanical insecticides, which employ plant extracts as active components, are less harmful to humans and the environment than synthetic pesticides (Dadang and Prijono, 2009). These plant-natural insecticides are employed directly by drying, grinding, and blending their powders with grains, or they are utilized indirectly by extracting dangerous chemicals in various methods and combining them with alcohol (Derbalah and Ahmed, 2011). These substances can affect the insect's nutritional behaviour, act as repellents or inhibitors, slow the insect's growth and development, or turn it infertile (Al-Joary *et al.*, 2021). Farmers use a variety of plant botanicals as insecticides, including Neem, garlic, onions, stinging nettle, Mexican sunflower, Chilies, Hot pepper, Tobacco, Pyrethrum, and Chilies (Kareru *et al.*, 2013). Citronella, eucalyptus, cinnamon, turmeric, ginger, and galangal might be used to protect stored products (Lee *et al.*, 2004), contact insecticides (Ngamo *et al.*, 2007),

repellent effects (García *et al.*, 2005), and also reduce ovicidal activities (Raja and William, 2008).

Very few studies are available in the Kurdistan region of Iraq about the comparative effects using some plant extracts of stinging nettle (*Urtica dioica*) to control (*Tribolium confusum*). Therefore, the purpose of this study to investigating the effective compounds of *U. dioica* extract on 3rd instar larvae of CFB and compare the effect of two different methods of extractions.

### 2. MATERIALS AND METHODS

#### 2.1 Insect rearing

The infected flour by *T. confusum* was obtained from Kurdistan shaking store for flour/Sumel region/Duhok/Iraq, then placed inside a container of 3L capacity and added to it 2kg of sterilized flour (stored in the freezer for one week). After one month, when the growth rate of CFB was increased they were placed in 10 wide-mouth plastic jars of 1L volume, 100 adult insects were placed, after covering the nozzles of the bottles with sterile gauze and fixing them with rubber bands, they were kept in an incubator in below conditions (Saleem *et al.*, 2013). The beetles were fed on flour containing 5% yeast. In an incubator, the plastic containers were kept in the dark. (Esmaili *et al.*, 2013). Rearing was controlled under incubator circumstances (Binder) ( $30 \pm 1^\circ\text{C}$  and  $70 \pm 5\%$  R.H) (Al-Sinjari and Al-Attar, 2015). This experiment utilized the 3<sup>rd</sup> instar larvae of *T. confusum*.

#### 2.2 Botanical insecticide

Nettle (*U. dioica* L.) is a Urticaceae family member that can be found in North America and Europe, and it is a perennial herb with stinging trichomes. (Bisht *et al.*, 2012). The plant trichomes are naturally built to defend themselves from insects (Kregiel *et al.*, 2018). Which contains formic acid, histamine,

\* Corresponding author

This is an open access under a CC BY-NC-SA 4.0 license (<https://creativecommons.org/licenses/by-nc-sa/4.0/>)

acetylcholine, and serotonin (Austin, 2008; Bourgeois et al., 2016).

### 2.3 Plant collection

*Urtica dioica* was collected from Derishke and Qumurey village/ Kani Massi sub-district/ Kurdistan Region/ Iraq in May 2021. After collection, the plants were washed (for removing the dust) with tap water, the plants were dried in shadow and the entire plant (except the root) was ground into fine powder by an electric mill cutter (Silver Crest 3000W), then the obtained powder was kept in closed plastic bags and placed at room temperature until the time of use (Abdulmajeed and Al-Chalabi, 2017).

### 2.4 Methods for the Plant Extraction

The extraction of bioactive chemicals from *U. dioica* has been developed utilizing both traditional procedures and various innovative techniques, including maceration, reflux, Soxhlet, hydro distillation, ultrasound- and microwave-assisted extraction (UAE), and microwave-assisted extraction (MAE). When using these methods to extract bioactive chemicals, ethanol was the most productive organic solvent used, leading to the greatest extraction yield. Additionally, the yield of isolated chemicals increased when extraction time was increased. The Soxhlet method provided the highest extraction yield among traditional techniques (Bandar et al., 2013).

Two methods of extraction were used in this study: Soxhlet and solvent extraction methods.

**2.4.1 Soxhlet apparatus:** The plant extract was prepared according to Banat et al. (2013) with slight variation. For one cycle of the Soxhlet apparatus, 20 gm of plant powder (*U. dioica*) was placed in the Soxhlet continuous extraction apparatus. The oil was extracted from it with 300 ml of ethanol 70% (60-80°C boiling point). The yielded solution was later filtered using filter papers and stored in a refrigerator until separated by a vacuum rotary evaporator.

**2.4.2 Solvent extraction method:** The plant extract was prepared according to Bashir and Al-Habib, (2021) with slight modification, where 50 gm of *U. dioica* powder had been mixed with 200 ml of ethanol 70% and put in a flask of 1000 ml capacity, and stored in ultra-low temperature freezer (-10 to -15 C°) for 24 hrs. Then, all solids were eliminated by a filtration process that requires just a vacuum pump, a Buchner funnel, two filter papers, and a vacuum flask. After the filtration (Whatman No.1 filter paper), a solution of ethanol and extract remained. Both methods of extraction need to be separated from the ethanol, for separating a rotary evaporator was used at 45 C°, 80 rpm, and 0 bar. The resultant extract or oil was fully ethanol-free (Scientific, 2018). The extracts were put in glass Petri plates and dried in an oven until the weight of the extracts was set before being stored in a refrigerator until needed.

### 2.5 Bioassay

Four concentrations were prepared from stock solution (0.5, 1, 1.5, 2.5 mg/ml) using dechlorinated tap water, and were used for spraying the 3<sup>rd</sup> instar larvae of *T. confusum*. Thirty larvae for each concentration are placed in sterile Petri dishes (100 X 15mm) divided into three sections (ten for each section) and enclosed by a Petri dish to avoid insects escaping. During the bio-test, each day the number of dead larvae was recorded. The larval mortalities were calculated and compared with the control (sprayed with dechlorinated water) of both methods. For each test, three replications were set up. Treated larvae in all groups were incubated at 30 ± 1 C° and R.H of 70% ± 5 (Al-Sinjari and Al-Attar, 2015). The Results were calculated after 24, 48, and 72 hours. The percentage of larval mortality was determined using the Abbott formula (Abbott, 1925).

$$M\% = \frac{M\% \text{ in certain dose} - \text{control } M\%}{(100) - \% \text{ control mortality}} * 100 \quad (1)$$

Where  $M$  = Mortality

### 2.6 Statistical Analysis

The data were analysed using the analysis of variance technique in the form of Two-way ANOVA, using the Minitab software package 17. Subsequently, Tukey's test was used to compare means.

### 2.7 Ethical approval

Ethical approval was obtained from the Protocol Review Committee of the Biological Sciences Committee (BSCZ) of the University of Zakho (ID: "BSCZ/25/7/2021").

## 3. RESULT AND DISCUSSION

### 3.1 The effect of *U. dioica* on the 3<sup>rd</sup> instar larvae using the Soxhlet method

The goal of this experiment was to evaluate the insecticidal activity of *U. dioica* extracts against *T. confusum* third-instar larvae. **Table (3.1)** shows the effectiveness of plant extracts on *T. confusum* mortality exposed to various doses across various periods by the Soxhlet extraction method.

The percentage of mortality increased as the dose of plant extract increased. The highest mortality rate was 100% at 1.5 mg/ml, while the lowest percentage of mortality 73.33% was observed after 24 hrs of exposure at 0.5 mg/ml dose.

**Table 3-1:** The effect of *U. dioica* extract on the 3<sup>rd</sup> instar larvae of *T. confusum* at different exposure periods by the Soxhlet method.

Doses mg/ml	% Mortality		
	24 hrs	48 hrs	72 hrs
Control	16.66 d	16.66 d	16.66 d
0.5	73.33 c	80 c	83.33 bc
1	96.66 ab	96.66 ab	96.66 ab
1.5	100 a	100 a	100 a
2.5	100 a	100 a	100 a
	Times	Dose	Times*Doses
P value	0.611	< 0.0001	0.879
F value	379.44	0.50	0.50

At a p-value of 0.05, the means that do not share a letter in any of the columns are statistically significant.

It was detected that there was a significant effect of doses on mortality ( $p < 0.001$ ), but the effect of time and interactions was found to be not significantly affected ( $p > 0.05$ ) mortality. The significantly lower value of mortality was recorded under control compared to another dose. Generally, the mortality increased with increasing doses. No significant interaction was found between time and dose, meaning that the effect of dose was the same for three periods of time.

### 3.2 The effect of *U. dioica* on the 3<sup>rd</sup> instar larvae using solvent extraction method

**Table (3.2)** displays the effect of *U. dioica* with different doses on the mortality percentage of 3<sup>rd</sup> instar larvae of *T. confusum*. The percentage of mortality increased as the dose of plant extract increased. Where the highest mortality rate was 1 of 00% at 2.5 mg/ml concentration, while the lowest percentage of mortality was 90% observed after 24 hrs of exposure at 0.5 mg/ml dose.

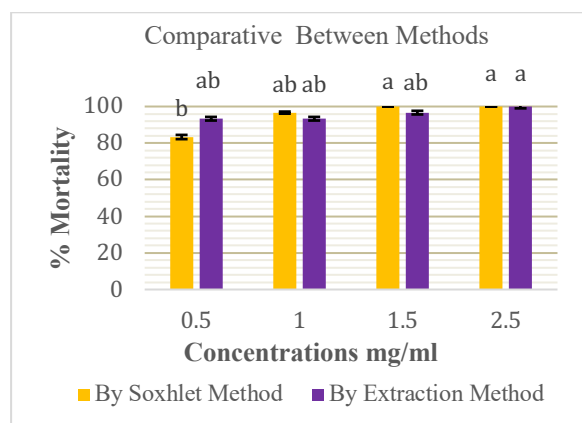
**Table 3-2:** The effect of *U. dioica* extract on the 3<sup>rd</sup> instar larvae of *T. confusum* at different exposure periods by Extraction method.

Doses mg/ml	% Mortality		
	24 hrs	48 hrs	72 hrs
Control	16.66 b	16.66 b	20 b
0.5	90 a	90 a	93.33 a
1	90 a	90 a	93.33 a
1.5	93.33 a	96.66 a	96.66 a
2.5	100 a	100 a	100 a
	<b>Times</b>	<b>Doses</b>	<b>Times*Doses</b>
<b>P value</b>	0.285	<0.0001	0.993
<b>F value</b>	497.68	1.30	0.22

The effect of dosages on mortality was found to be significant ( $P < 0.001$ ), however, the effect of time and interactions with dose on mortality was not significant ( $p > 0.05$ ). The significantly lower value of mortality was recorded under control compared to other doses. By raising the dosage, the mortality rate was increased. Interactions between time and dose were determined to be non-significant, indicating that the dose had the same effect over three time periods.

### 3.3 The comparative effect of *U. dioica* on the 3<sup>rd</sup> instar larvae using Soxhlet and solvent extraction methods

Figure (3.1) shows the comparative effect of treatment with different concentrations and two methods of extractions on the mortality percentage of 3<sup>rd</sup> instar larvae of *T. confusum*.



**Figure 3-1:** The comparative effect of *U. dioica* extract on the 3<sup>rd</sup> instar larvae of *T. confusum* after 72 hrs exposure by using Soxhlet and Extraction methods.

The results revealed that there were no significant differences between both used methods in terms of their effect on the mortality of insects ( $P > 0.05$  and  $F = 0.13$ ). In general, increasing doses led to a rise in mortality. Regarding interactions, no significant interaction was seen between the method and dose.

The mortality of the larvae via both procedures may be attributable to the presence of chemical compounds such as formic acid (Coile, 1999). Lectin and iron in great quantity, histamine, and serotonin (Austin, 2008; Bourgeois et al., 2016; Kregiel et al., 2018). These compounds might disturb the digestive system of larvae especially the midgut and gastric caecum (Ndione et al., 2007). Furthermore, extremely hazardous plant extracts are rich in non-polar saturated fatty acids that rapidly permeate the cuticle layers and subsequently have a harmful effect. (Zhou et al., 2005). In addition to the persistence of extracts on the outside surface of the insect's body, which covers the spiracles and so impedes the insect's ability to breathe, the extracts also close the insect's spiracles (Fathi and Shakarami, 2014). James et al. (2015) Researchers

examined the relationship between parasitic, predatory, and helpful insects with *U. dioica*. The research concluded that stinging nettle may serve as an essential home for helpful insects and play a role in encouraging biological management to protect crops.

These results are consistent with prior studies indicating that several plant extracts are poisonous to *T. confusum* larvae. Numerous studies have shown that some plant extracts have potent larvicidal actions against insect pests (Liu et al., 2012; Mahfuz and Khanam, 2007; Zaka et al., 2019). The results of the current research were similar to the study of sixteen aromatic plant extracts that were assessed for their contact toxicity on *T. confusum* larvae. *Mantisalca duriaei* methanolic and ethyl acetate extracts and *Rhaponticum acaule* petroleum ether, chloroformic, and methanolic extracts substantially inhibited larval growth. Although only *R. acaule* has a very toxic impact on larvae for all extracts, larval mortality was the greatest. Using petroleum ether and methanol extracts of *R. acaule*, it attained 83% and 77%, respectively (Boussaada et al., 2008). The results of the current study are relatively in accordance with those reported by Ali and Mohammed (2013) who investigated the lethality of the larval stage of *T. confusum* using methanol extracts of six indigenous plants from the Kurdistan area. The most effective of the six test plant extracts was *Zingiber officinale* with a 100% death rate at 2 hours, followed by *Anethum graveolens* at 56.67% after 4.5 hours, which is the least poisonous to larvae.

In addition, the comparable study investigated the repellence of ethanolic extracts of *E. glauca* against *T. confusum* larvae. Effective repellents have been discovered in all plant extracts, with the *E. glauca* activating at a concentration of 7.5% to 100% repellency (Mohammed, 2013).

A study was accepted to assess the toxicity of lambda-cyhalothrin and the essential oils of cardamom seeds from *Elettaria cardamomum* L., either separately or in combination, to the larval stages of the *T. confusum*. The outcomes demonstrated that the oil extract was substantially more effective than the insecticide lambda. Larvae exposed to cardamom oil at concentrations of 0.1, 0.3, 0.5, and 1 µl/larva experienced mortality rates of 60, 76.67, 90, and 96.67 percent, respectively (Al-Sinjari and Al-Attar, 2015). The effect of cardamom seeds extracts was the same as the effect of Nettle extracts used in this research. This is consistent with the results of Saidana et al. (2007), who discovered that petroleum ether extracts from a variety of plant species were very efficient against CFB larvae and adults, with the larvae being more susceptible than the adults. Another study by AL-Jamil and AL-Sinjari (2019) showed the death rate of *T. confusum* increased as the ratio of castor extract to insecticide lambda increased, and the toxicity varied depending on the surface type.

Also, Khoshnoud and Khayami (2008) addressing the insecticidal activities of an ethanolic extract of *Verbascum cheiranthifolium* against two insect pest species of stored products demonstrated that insect mortality rose as the dose rate increased. AL-Attar (2006) indicates that Actara has strong insecticidal action and would be very successful in controlling *T. confusum*. Fathi and Shakarami (2014) analysis of essential oils from five species of Eucalyptus on *T. confusum* larvae for fumigant toxicity. The results suggested that *E. viminalis* essential oils were the most harmful to CFB larvae. In another experiment, Khanam et al. (1990) stated that seed coat extracts of *Aphanamixis polystachea* had a lethal effect on the growth and development of *T. confusum*.

## 4. CONCLUSION

The toxicity of *Urtica dioica* extracts increases as concentrations rise for both methods of extractions, but there were no significant differences between both methods,

meaning that both methods had a similar effect on insect's larvae. The effect of periods of time also had statistically the same effect at the first time of exposure, meaning that the effect of plant extract was lethal after the first period.

## 5. ACKNOWLEDGEMENTS

I would like to thank and appreciate my supervisors from the bottom of my heart, Prof. Dr. Badia, an M. Al-Chalabi, and Assist. Prof. Dr. Sofyan Haje Sedo, who helped me finish this work, for their notes on how to work and write the project, for their guidance throughout this study, and for their patience, support, and encouragement. My heartfelt thanks go to Dr. Ahmad Basheer, head of the Biology Department/at the University of Zakho, for his generous assistance in completing this study.

I am grateful to Dr. Saleem E. Shahbaz of Duhok University helped me classify the plant species utilized in this study, and Dr. Razaq Shealan Akl of the Iraqi National History Research Center and Museum helped me categorize the insect species used in this study. And also, Mr. Hajar A. Ameen, College of Agricultural Engineering Sciences, University of Duhok, for his assistance with the statistical analysis of this study's data. Last, of all, I'd like to express my appreciation to Mr. Thamer Mohammed for his assistance during my studies at Animal house.

## REFERENCES

- Abbott, W. S. (1925). A method of computing the effectiveness of an insecticide. *J. Econ. Entomol.*, 18(2), 265–267.
- Abdulmajeed, A. M., & Al-Chalabi, B. (2017). The Cumulative Effects of Plant Extract *Tavri Shrub* (*Daphne mucronata*) on Some Biological Aspects of Mosquito *Culex molestus* Forskal. *Science Journal of University of Zakho*, 5(2), 167–171.
- AL-Attar, H. J. (2006). Comparison of toxicity and residual effectiveness of Actara®(Thiamethoxam®) with three pyrethroid insecticides against five Coleopteran species infesting stored-products. *Tikrit Journal of Pure Science*, 11(2).
- AL-Jamil, S. K., & AL-Sinjari, S. H. S. (2019). The Effect of the Extent of the Exposure and Concentration for the Three Types of Surface Treatment in the Mortality of Larvae and Adults of Confused Flour Beetle, *Tribolium confusum* (Du Val). *Tikrit Journal for Agricultural Sciences*, 18(3), 100–107.
- Al-Joary, Y. I. M. A., Al-Obaidi, R. G., & Al-Ejaidy, S. (2021). 'study of the repellent activity of some medicinal herbs powder against adults of *Tribolium confusum* duv.(tenebrionidae: coleoptera).' *Plant Archives*, 21(1), 445–450.
- Al-Sinjari, S. H. S., & Al-Attar, H. J. (2015). Toxic effects of essential oils of *Elettaria cardamomum* L. and *Lambda-Cyhalothrin* on *Tribolium confusum* (Duval). *SJUOZ*, 3, 15–26.
- Al Abadi A. Q. et al. (2008). The attractiveness and repellency effect of some vegetable oils in the adult confused flour beetle. *Tribolium confusum* duv.(Tenebrionidae, Coleoptera)', journal of technique, 21(2). *Journal of Technique*, 21(2).
- Ali, W. K., & Mohammed, H. H. (2013). Toxic effect of some plant extracts on the mortality of flour beetle *Tribolium confusum* (Duval)(Coleoptera: Tenebrionidae). *Entomol Ornithol Herpetol*, 2(115), 983–2161.
- Austin, S. B. (2008). *The Paradox of L., "Stinging Nettle": A Blessing and a Curse*.
- Banat, F., Pal, P., Jwaied, N., & Al-Rabadi, A. (2013). Extraction of olive oil from olive cake using Soxhlet apparatus. *American Journal of Oil and Chemical Technologies*, 1(4), 1–8.
- Bandar, H., Hijazi, A., Rammal, H., Hachem, A., Saad, Z., & Badran, B. (2013). Techniques for the extraction of bioactive compounds from Lebanese *Urtica dioica*. *American Journal of Phytomedicine and Clinical Therapeutics*, 1(6), 507–513.
- Bashir, T. M., & Al-Habib, O. A. M. (2021). The Role of K<sup>+</sup>, Ca<sup>2+</sup> channels and Endothelial Hyperpolarizing Factors in Vasorelaxation Induced by *Tribulus terrestris*. *Technium BioChemMed*, 2(1), 94–100.
- Bisht, S., Bhandari, S., & Bisht, N. S. (2012). *Urtica dioica* (L): an undervalued, economically important plant. *Agric Sci Res J*, 2(5), 250–252.
- Bourgeois, C., Leclerc, É. A., Corbin, C., Doussot, J., Serrano, V., Vanier, J.-R., Seigneuret, J.-M., Auguin, D., Pichon, C., & Lainé, É. (2016). Nettle (*Urtica dioica* L.) as a source of antioxidant and anti-aging phytochemicals for cosmetic applications. *Comptes Rendus Chimie*, 19(9), 1090–1100.
- Boussaada, O., Kamel, M. B. H., Ammar, S., Haouas, D., Mighri, Z., & Helal, A. N. (2008). Insecticidal activity of some Asteraceae plant extracts against *Tribolium confusum*. *Bulletin of Insectology*, 61(2), 283–289.
- Coile, N. C. (1999). *Urtica chamaedryoides Pursh: a stinging nettle, or fireweed and some related species*. Fla. Department Agric. & Consumer Services, Division of Plant Industry.
- Dadang, E. D. F., & Prijono, D. (2009). Effectiveness of two botanical insecticide formulations to two major cabbage insect pests on field application. *J. ISSAAS*, 15(1), 42–51.
- Derbalah, A., & Ahmed, S. (2011). Oil and powder of spearmint as an alternative to *Sitophilus oryzae* chemical control of wheat grains. *Journal of Plant Protection Research*.
- Esmaili, M., Vojoudi, S., & Parsaeyan, E. (2013). Fumigant toxicity of essential oils of *Mentha pulegium* L. on adults of *Callosobruchus maculatus*, *Tribolium castaneum*, *Lasioderma serricorne* and *Sitophilus oryzae* in laboratory conditions. *Technical Journal of Engineering and Applied Sciences*, Giza, 3(9), 732–735.
- Fathi, A., & Shakarami, J. (2014). Larvicidal effects of essential oils of five species of *Eucalyptus* against *Tribolium confusum* (du Val) and *T. castaneum* (Herbst). *International Journal of Agriculture and Crop Sciences (IJACS)*, 7(5), 220–224.
- García, M., Donadel, O. J., Ardanaz, C. E., Tonn, C. E., & Sosa, M. E. (2005). Toxic and repellent effects of *Baccharis salicifolia* essential oil on *Tribolium castaneum*. *Pest Management Science: Formerly Pesticide Science*, 61(6), 612–618.
- James, D. G., Lauby, G., Seymour, L., & Buckley, K. (2015). Beneficial insects associated with stinging nettle, *Urtica dioica* Linnaeus, in central Washington State. *The Pan-Pacific Entomologist*, 91(1), 82–90.
- Kareru, P., Kipkorir, Z. R., & Wamaitha, M. E. (2013). Use of botanicals and safer insecticides designed in controlling insects: the African case. *Insecticides-Development of Safer and More Effective Technologies*, 10, 297–309.
- Khanam, L. A. M., Talukder, D., Khan, A. R., & Rahman, S. M. (1990). Insecticidal properties of *Roynia, Aphanamixis polystachya* Wall.(Parker)(Meliaceae) against *Tribolium confusum* Duval. *Journal of Asiatic Society of Bangladesh Science*, 16, 71–74.
- Khoshnoud, H., & Khayami, M. (2008). Insecticidal effects of ethanolic extract from *Verbascum cheiranthifolium* Boiss. against two stored-product insect pests species. *Journal of Biological Sciences*, 8(1), 191–195.
- Kregiel, D., Pawlikowska, E., & Antolak, H. (2018). *Urtica* spp.: Ordinary plants with extraordinary properties. *Molecules*, 23(7), 1664.
- Lee, B.-H., Annis, P. C., & Choi, W.-S. (2004). Fumigant toxicity of essential oils from the Myrtaceae family and 1, 8-cineole against 3 major stored-grain insects. *Journal of Stored Products Research*, 40(5), 553–564.
- Liu, Z. L., Zhao, N. N., Liu, C. M., Zhou, L., & Du, S. S. (2012). Identification of insecticidal constituents of the essential oil of *Curcuma wenyujin* rhizomes active against *Liposcelis bostrychophila* Badonnel. *Molecules*, 17(10), 12049–12060.
- Mahfuz, I., & Khanam, L. A. M. (2007). Toxicity of some indigenous plant extracts against *Tribolium confusum* Duval. *Journal of Bio-Science*, 15, 133–138.
- Mohammed, H. H. (2013). Repellency of Ethanolic Extract of Some Indigenous Plants against *Tribolium confusum*(Duval)(Coleoptera: Tenebrionidae). *Agric. Vet. Sci*, 2(6), 27–31.
- Ndione, R. D., Faye, O., Ndiaye, M., Dieye, A., & Afoutou, J. M. (2007). Toxic effects of neem products (*Azadirachta indica* A. Juss) on *Aedes aegypti* Linnaeus 1762 larvae. *African Journal of Biotechnology*, 6(24).
- Ngamo, T. S., Ngatanko, I., Ngassoum, M. B., Mapongmestsem, P. M., & Hance, T. (2007). Persistence of insecticidal activities of crude essential oils of three aromatic plants towards four major stored product insect pests. *African Journal of Agricultural Research*, 2(4), 173–177.
- Phillips, T. W., & Throne, J. E. (2010). Biorational approaches to managing stored-product insects. *Annual Review of Entomology*, 55, 375–397.

- Raja, M., & William, S. J. (2008). Impact of volatile oils of plants against the cowpea beetle *Callosobruchus maculatus* (FAB.) (Coleoptera: Bruchidae). *International Journal of Integrative Biology*, 2(1), 62–64.
- Saidana, D., Halima-Kamel, M. Ben, Mahjoub, M. A., Haouas, D., Mighri, Z., & Helal, A. N. (2007). Insecticidal activities of Tunisian halophytic plant extracts against larvae and adults of *Tribolium confusum*. *Tropicultura*, 25(4), 193.
- Saleem, M., Hussain, D., Rashid, R. S., Saleem, H. M., Ghouse, G. H., & Abbas, M. (2013). Insecticidal activities of two citrus oils against *Tribolium castaneum* (herbst). *American Journal of Research Communication*, 1(6), 67–74.
- Scientific, A. (2018). *Solvent extraction method of plant using ethanol*. [coleparmer.co.uk/blog/2018/06/28/steps-safe-solvent-extraction/](http://coleparmer.co.uk/blog/2018/06/28/steps-safe-solvent-extraction/)
- Verheggen, F., Ryne, C., Olsson, P.-O., Arnaud, L., Lognay, G., Högberg, H.-E., Persson, D., Haubruge, E., & Löfstedt, C. (2007). Electrophysiological and behavioral activity of secondary metabolites in the confused flour beetle, *Tribolium confusum*. *Journal of Chemical Ecology*, 33(3), 525–539.
- Zaka, S. M., Iqbal, N., Saeed, Q., Akrem, A., Batool, M., Khan, A. A., Anwar, A., Bibi, M., Azeem, S., & Bibi, R. (2019). Toxic effects of some insecticides, herbicides, and plant essential oils against *Tribolium confusum* Jacquelin du Val (Insecta: Coleoptera: Tenebrionidae). *Saudi Journal of Biological Sciences*, 26(7), 1767–1771.
- Zhou, S., Chan, S. Y., Goh, B. C., Chan, E., Duan, W., Huang, M., & McLeod, H. L. (2005). Mechanism-based inhibition of cytochrome P450 3A4 by therapeutic drugs. *Clinical Pharmacokinetics*, 44(3), 279–304.